

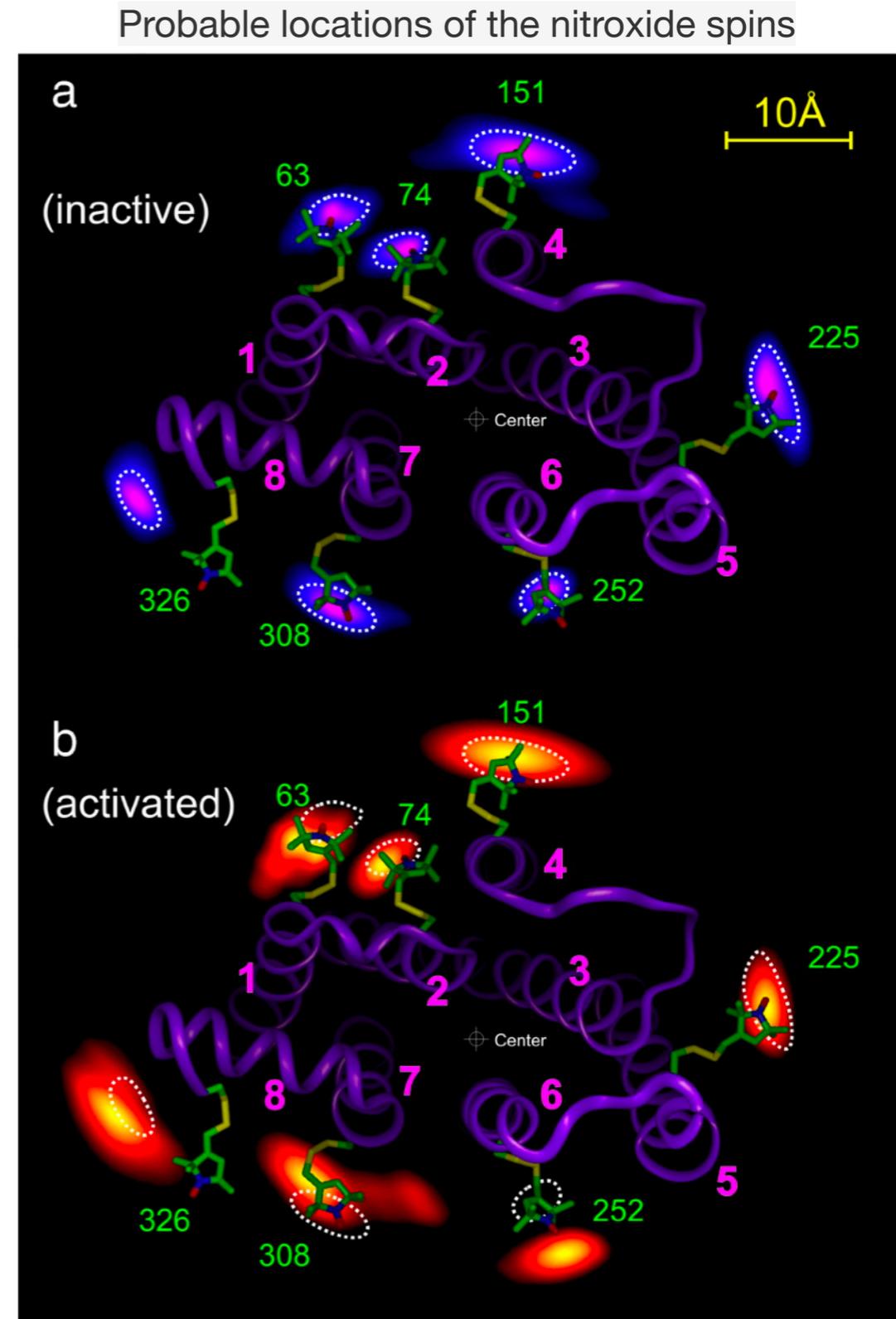
Theory and applications

MD simulations and other methods

Methods to measure the 'dynamics'

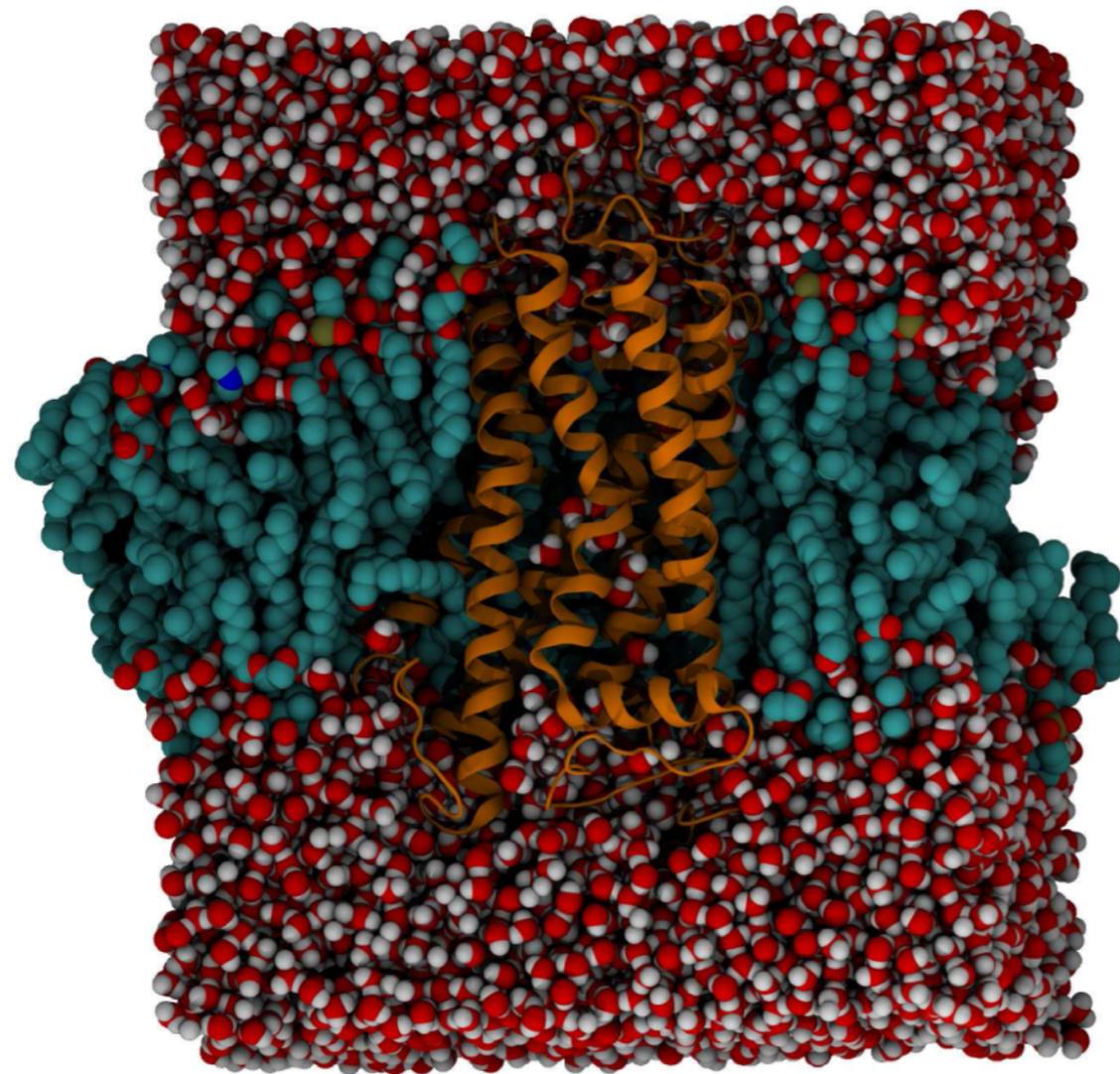
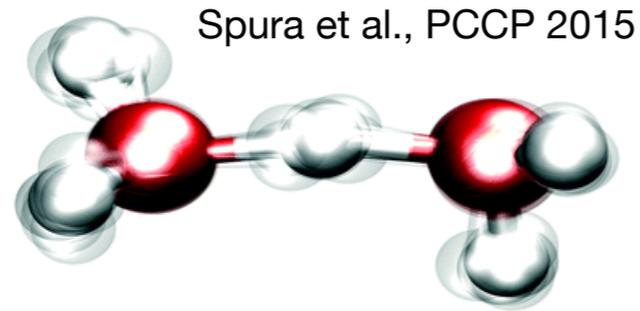
- NMR
- Labelling: EPR, DEER
- Labelling: single molecule FRET
- Raman, IR spectroscopy
- X-ray

- MD: provides the time resolved motion of atoms at atomic resolution



Altenbach et al., PNAS, 2008

MD simulations: what is it good for?



- **replace experiments:**
e.g. coupled clusters (only < 15 atoms) more accurate than experiments!
- **explain experiments:**
EPR, NMR, mutations, ...
- **drive experiments:**
by proposing hypothesis
- Generally promotes **interdisciplinary research**

MD simulations: pro and contra

- + time resolved physical movements of atoms and molecules
- + the dynamic evolution of the system —> ,trajectory‘
- + study the motions of biological macromolecules

- Only potential energy is optimized, not the free energy
- entropic contributions to thermodynamic stability neglected
- Only crude approximation of electrostatics in all atom force field

How to calculate interaction energies?

Calculate to what path a given physical system will take over time:

Quantum Mechanics (describes physical laws of nature on *an atomic scale*)

Schrödinger's equation: $H \Psi = E \Psi$ (time independent)

H = Hamiltonian (corresponds to total systems energy)

E = Energy Eigenvalue

Ψ = wavefunction

Classical Mechanics (describes motion of *macroscopic objects*):

Newton's second law of motion:

$$\mathbf{F} = m\mathbf{a}$$

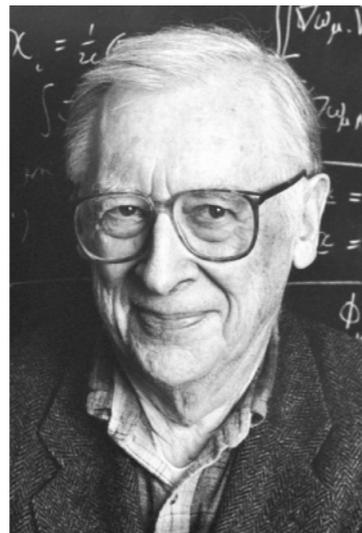
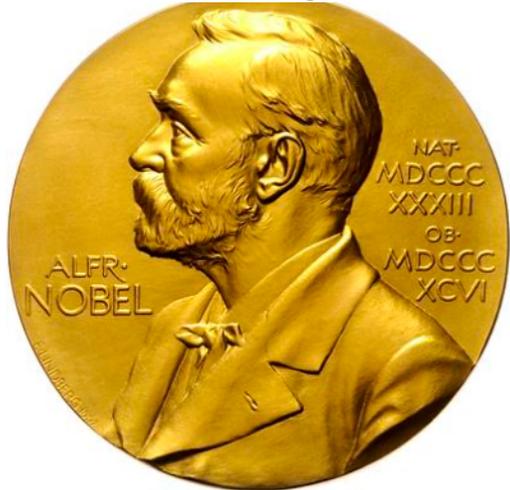
F = force

m = mass

a = acceleration

MD is established

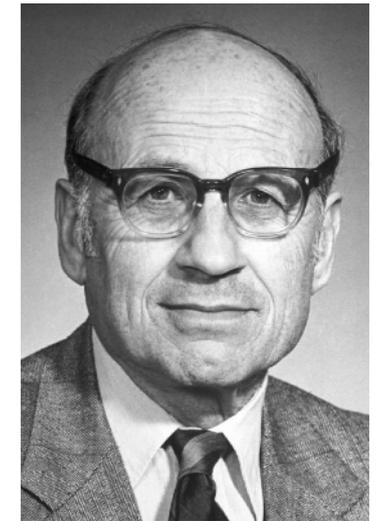
Chemistry 1998



John Pople

-John Pople "for his development of **computational methods in quantum chemistry**"

- Walther Kohn "for his development of the **density-functional theory**"



Walther Kohn

"for the development of **multiscale models for complex chemical systems**"

Chemistry 2013



Martin Karplus

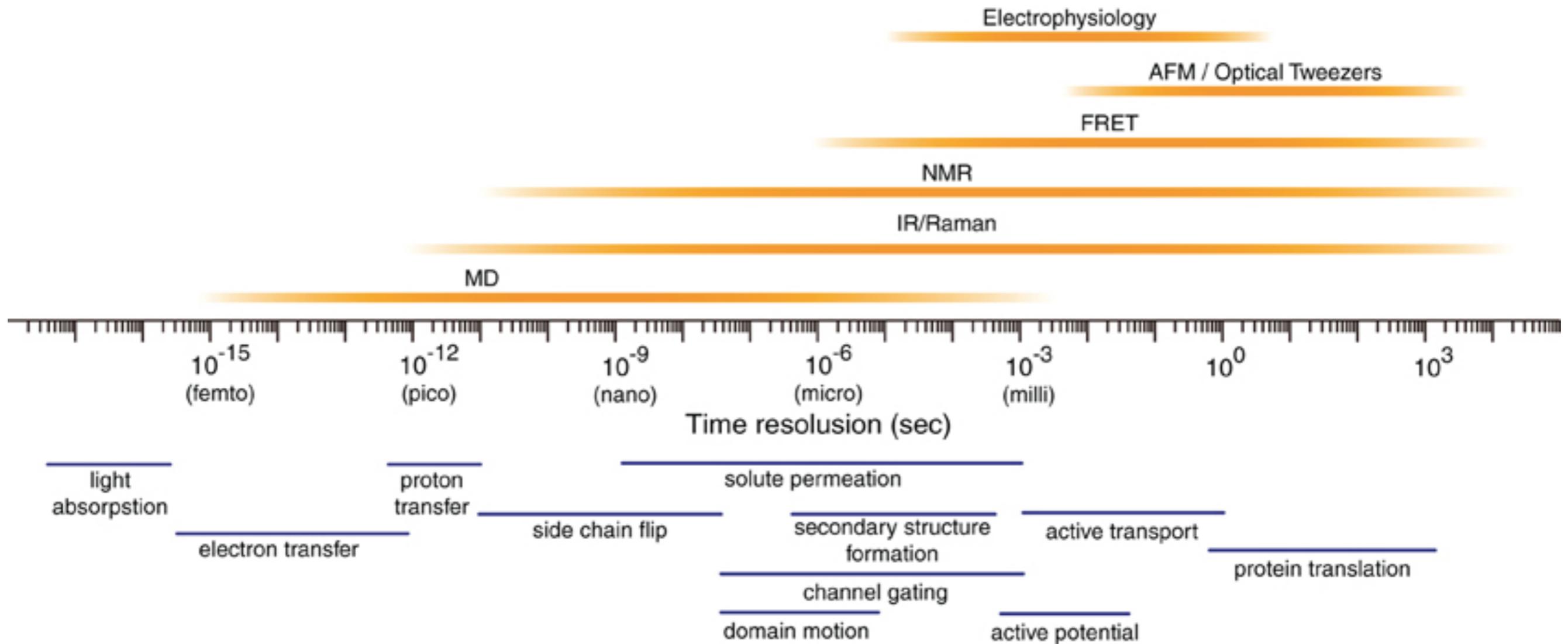


Michael Levitt

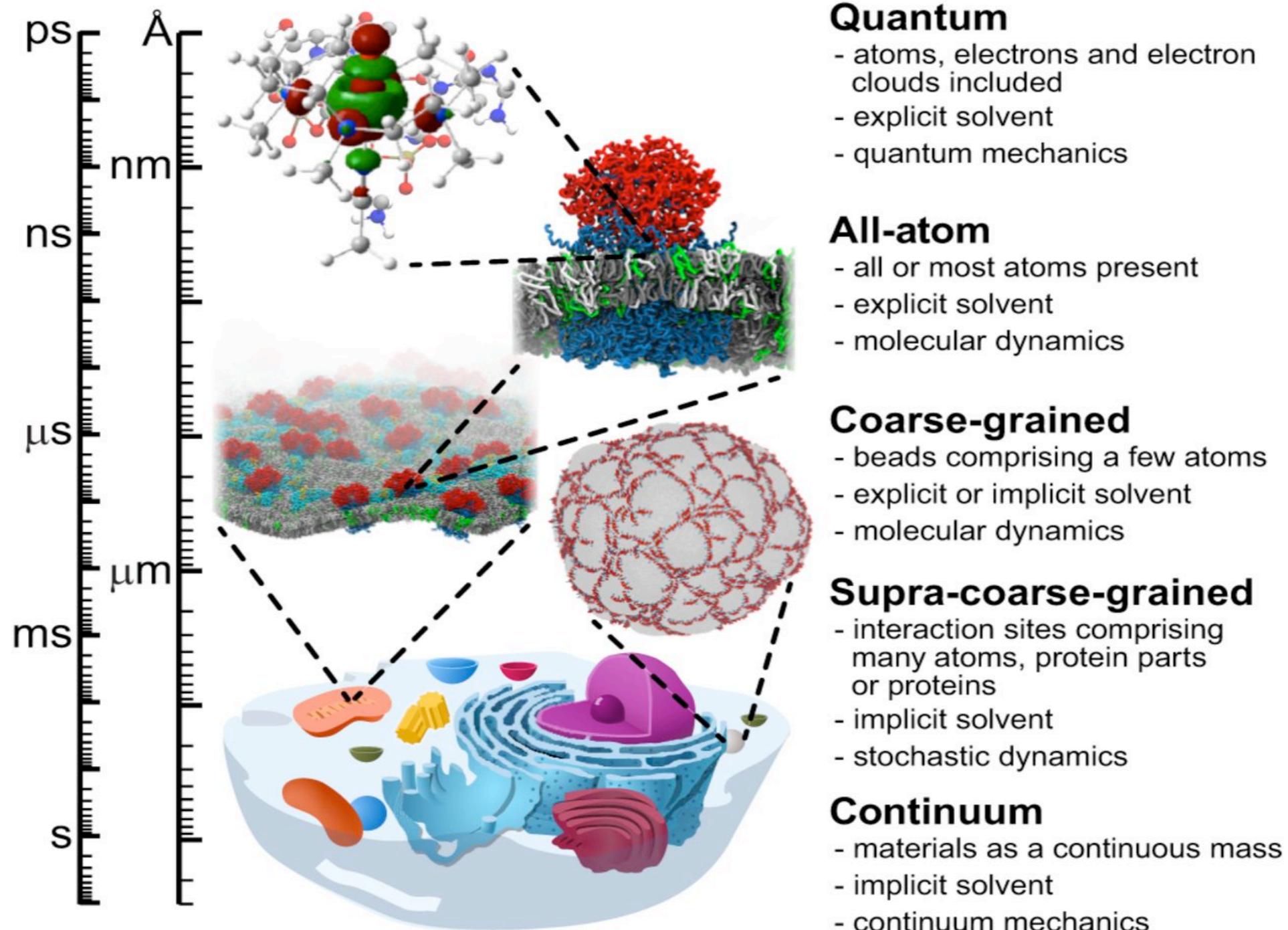


Arieh Warshel

MD simulations: Timescales



types of MD simulations



Computational 'microscopy' of cellular membranes Helgi I. Ingólfsson, Clément Arnarez, Xavier Periole, Siewert J. Marrink Journal of Cell Science 2016 129: 257-268; doi: 10.1242/jcs.176040

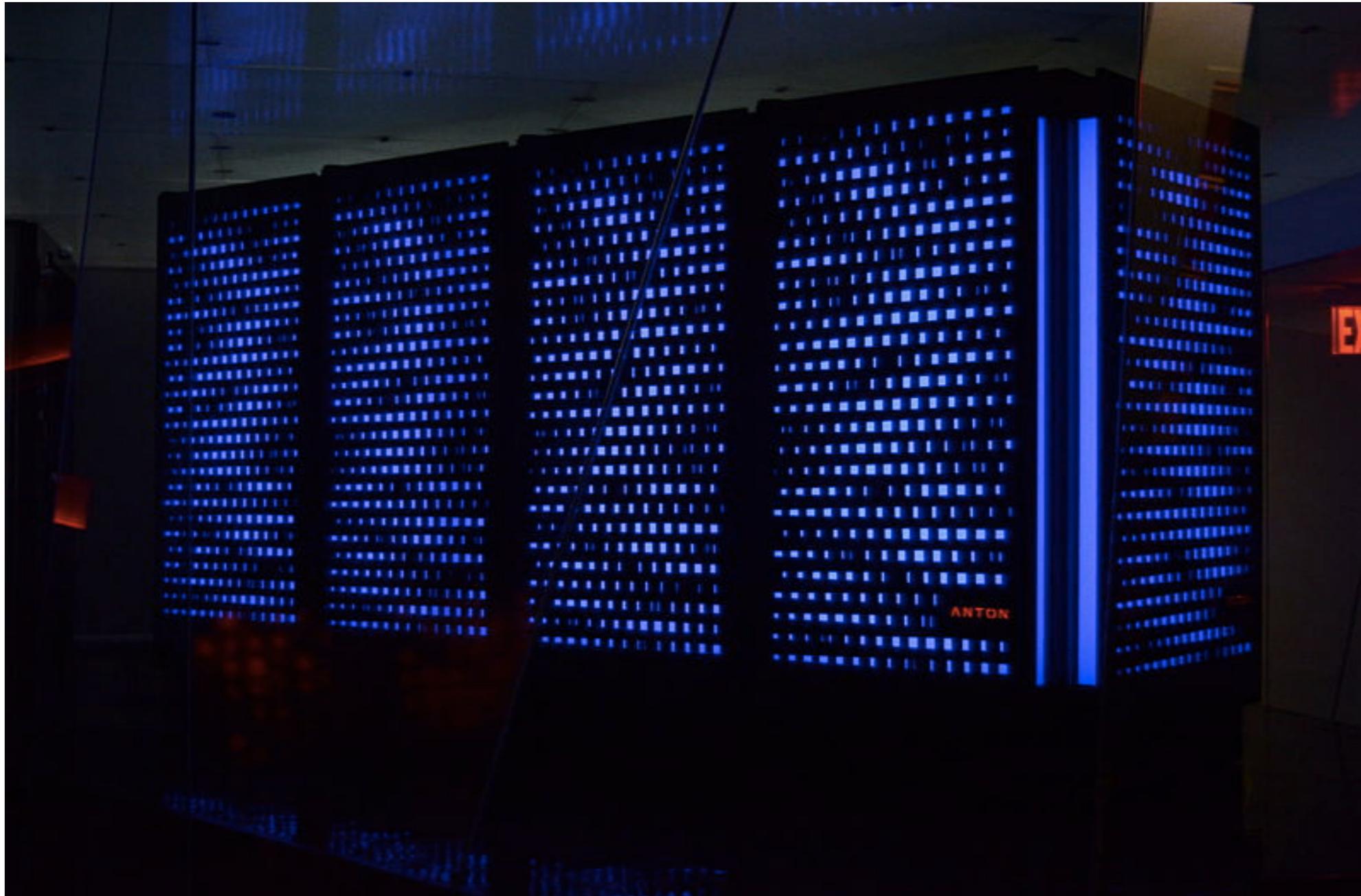
MD Simulations in a nutshell

Methods/Force Fields/Packages

- Molecular Mechanics: empirical force fields: AMBER, OPLS, CHARMM, GROMOS, ...
- Molekular Dynamics Packages (classical Newton-mechanics: GROMACS, NAMD...)
- Semi-empirical Molecular-Orbital-Theorie (MNDO, AM1, PM3, OM2, MNDO/d, ...)
- DFT (B3LYP, PW91, BP86, LDA...)
- *ab Initio* Molecular-Orbital-Theorie (Hartree-Fock, Møller-Plesset, Coupled Cluster ...)

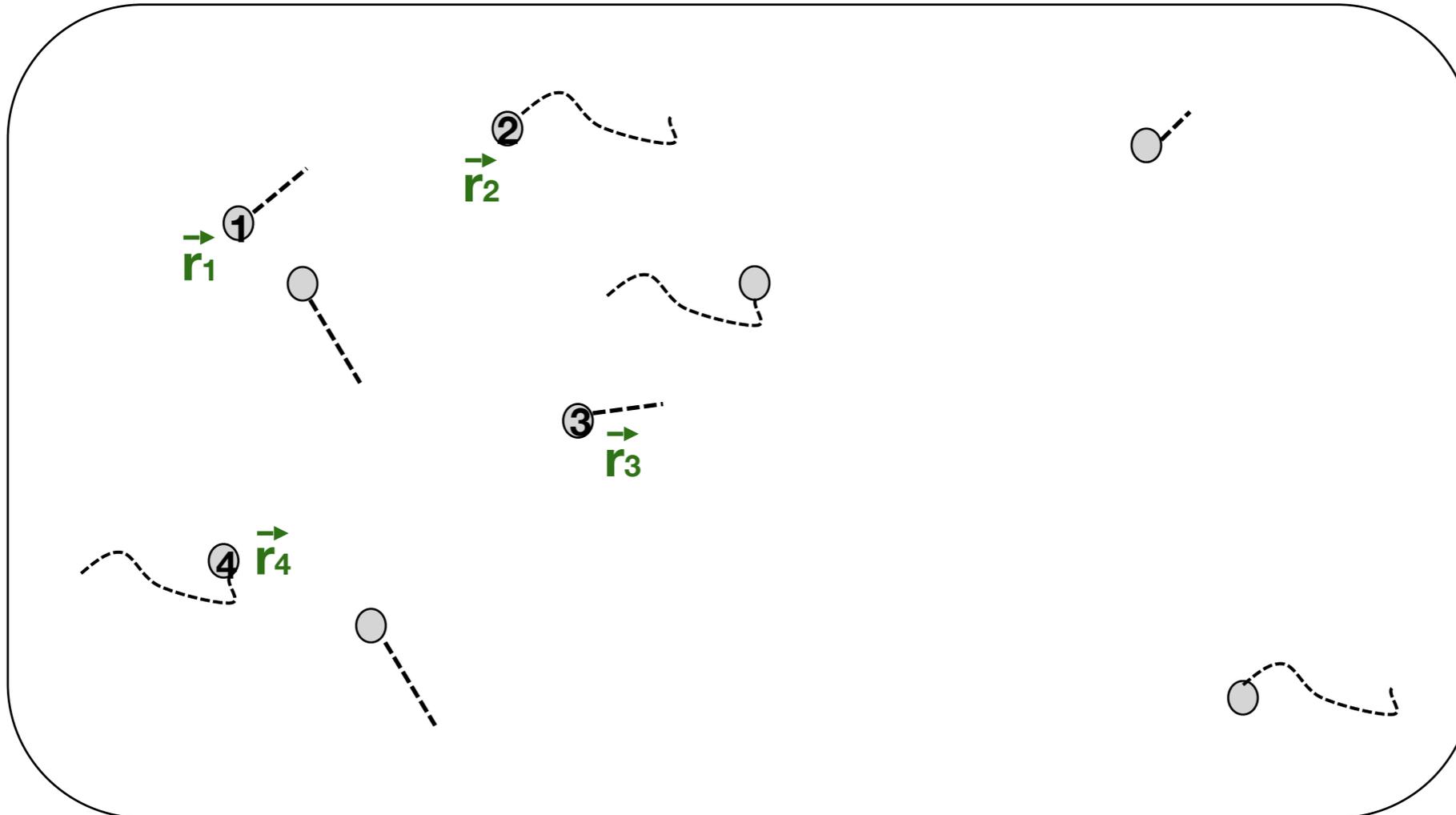
MD simulations: high computing resources

Anton at DE SHAW



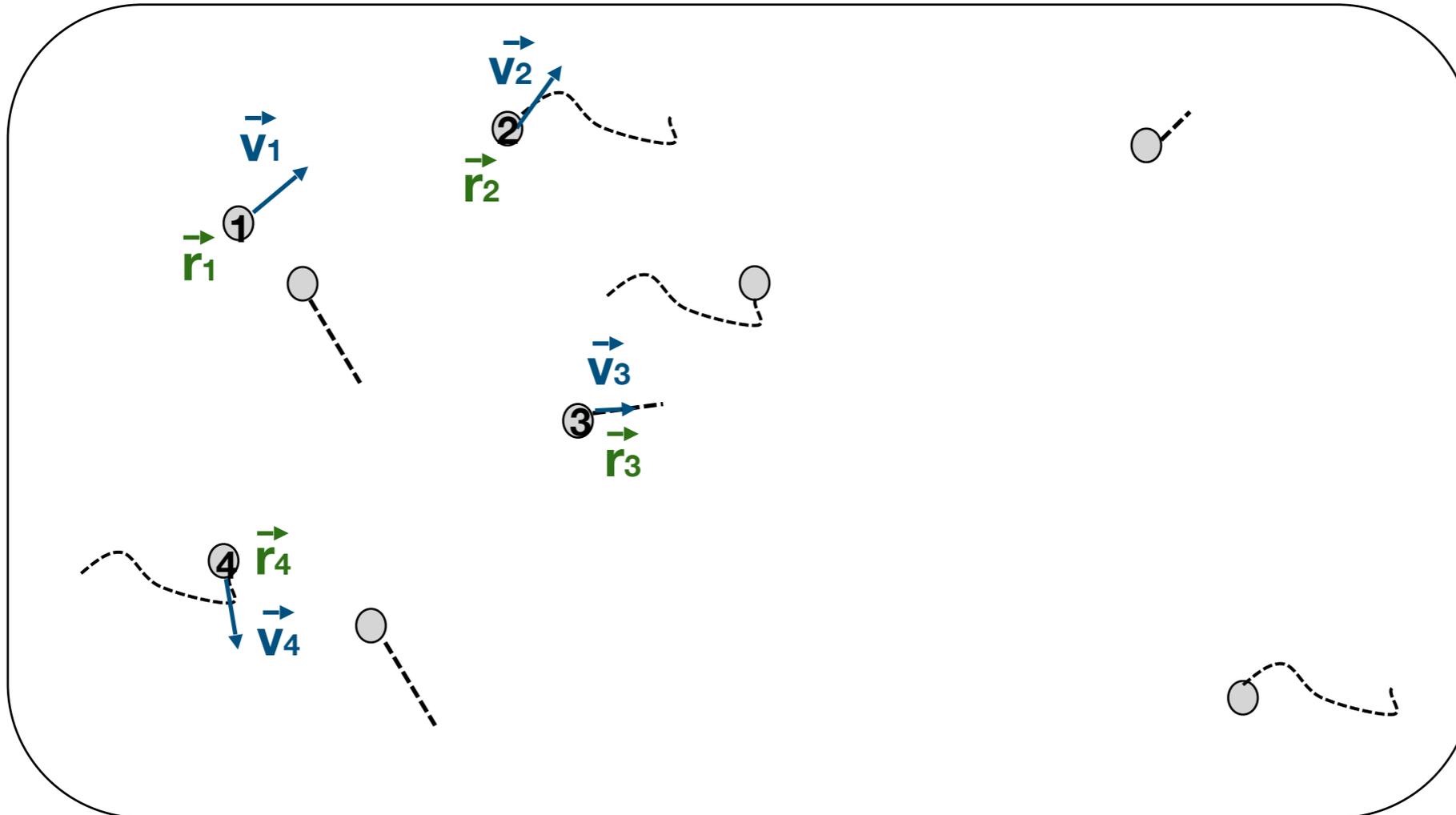
goal: predict motion of atoms

many-body system



- - Atom
- - motion
- \vec{r}_{1-n} - Initial position

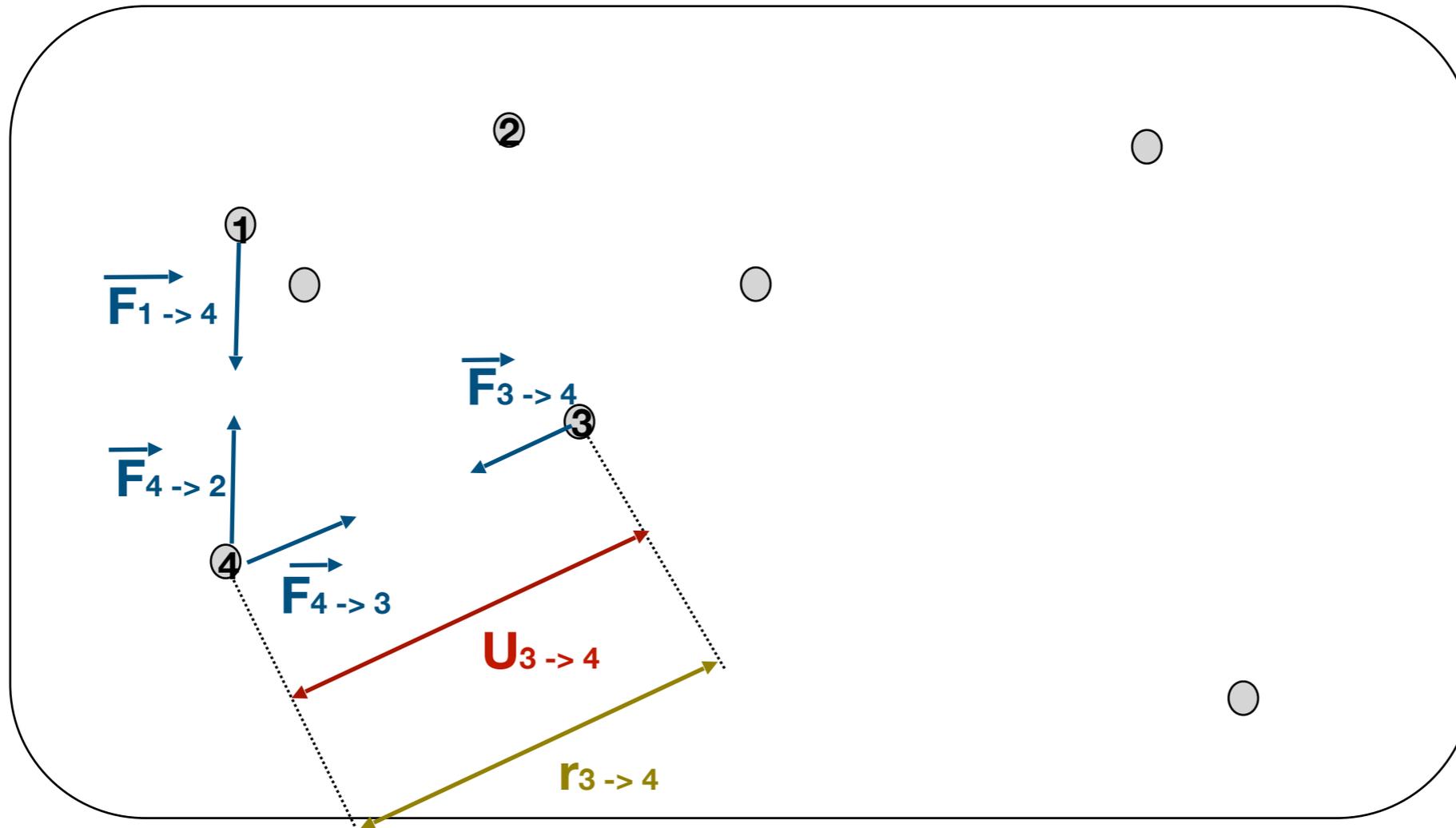
goal: predict motion of atoms



- - Atom
- motion
- \vec{r}_{1-n} - Initial position
- \vec{v}_{1-n} - Initial velocity

We have to calculate interaction energies

Interaction energies are approximated by the potential energy **U**



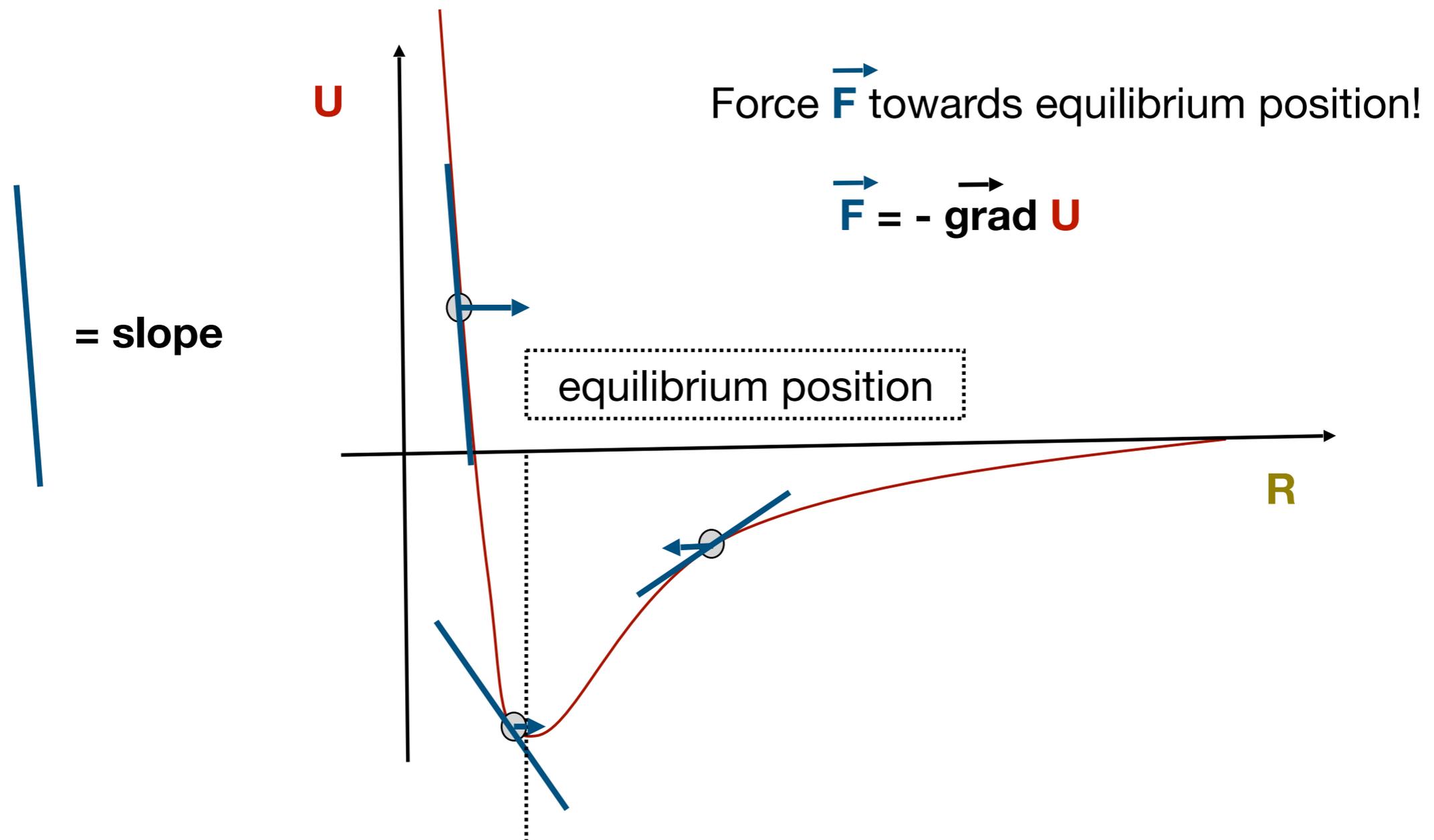
\vec{F} - Force of atom

U - potential energy

r - distance between atoms

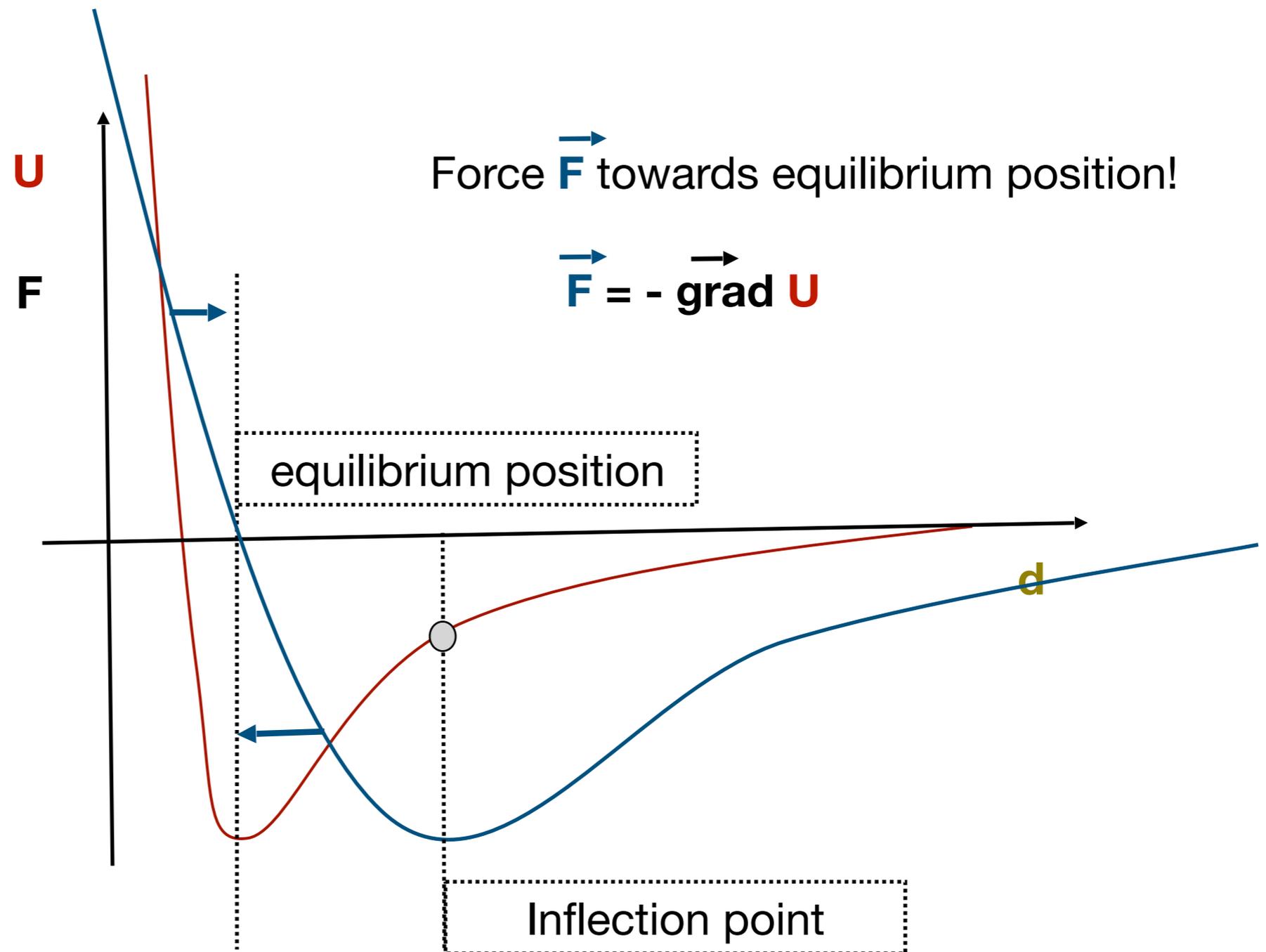
How to calculate interaction energies?

Approximate the force \vec{F} from the potential Energy U



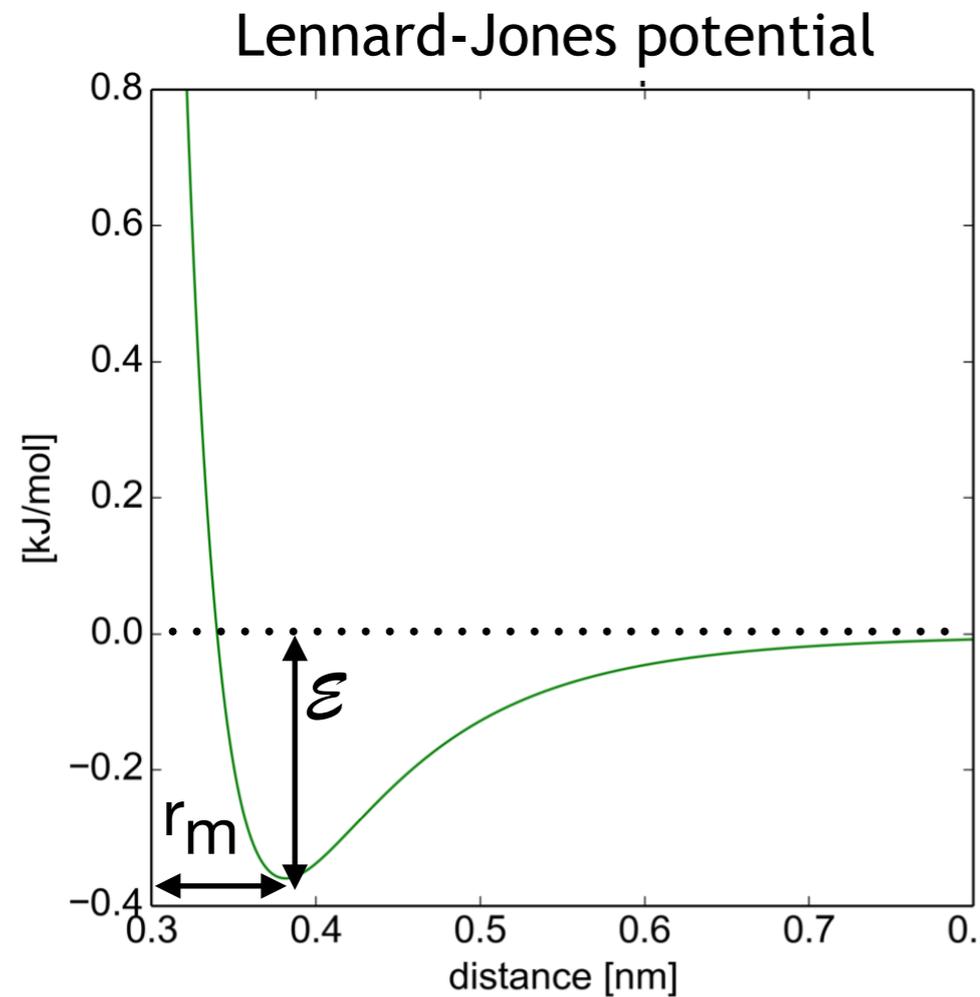
How to calculate interaction energies?

Approximate the force \vec{F} from the potential Energy U



Non-bonded interactions

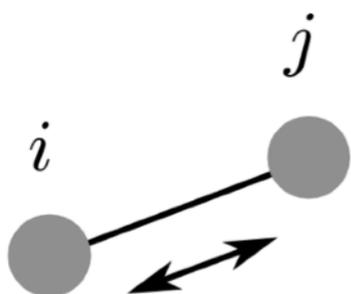
Interaction potential energies approximated by



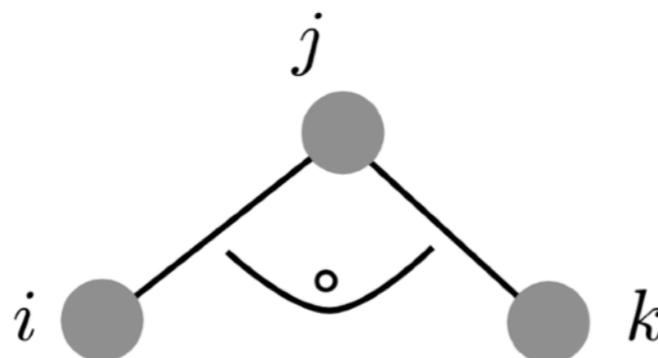
$$U_{vdw} = \sum_{i < j}^{atoms} \epsilon_{ij} \left[\left(\frac{r_m}{r_{ij}} \right)^{12} - 2 \left(\frac{r_m}{r_{ij}} \right)^6 \right]$$

Bonded interactions

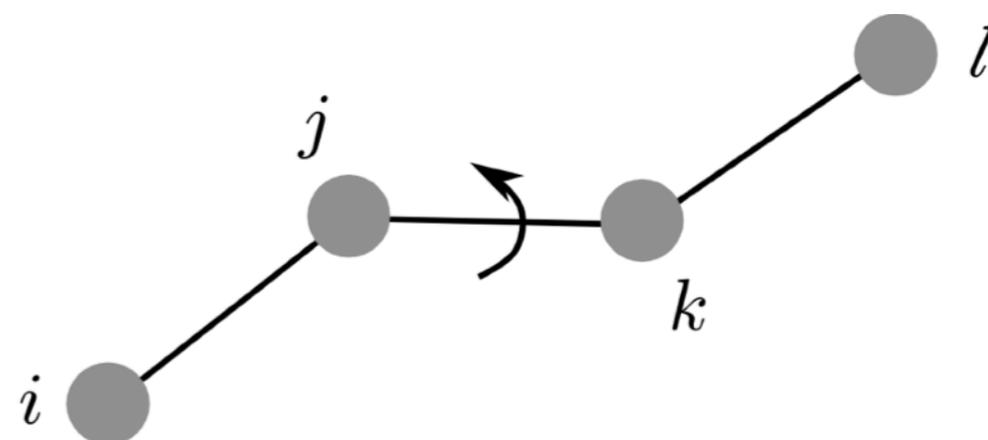
Bond lengths



Bond angles



Torsion angles



$$U_{angles} = \sum_{angles} K_{\theta} (\theta - \theta_{eq})^2$$

$$U_{Bonds} = \sum_{bonds} K_r (r - r_{eq})^2$$

$$U_{dihedrals} = \sum_{dihedrals} K_{\phi} (1 + \cos[n\phi - \gamma])^2$$

Allen, NIC 2004

Complete term of potential energy U

bonded

$$U(r^N) = \left(\left(\sum_{\text{bonds}} K_r (r - req)^2 \right) + \left(\sum_{\text{angles}} K_\theta (\theta - \theta_{eq})^2 \right) + \left(\sum_{\text{dihedrals}} K_\phi (1 + \cos[n\phi - \gamma])^2 \right) \right) + \sum_{i=1}^N \sum_{j=i+1}^N \epsilon_{ij} \left[\left(\frac{r_m}{r_{ij}} \right)^{12} - 2 \left(\frac{r_m}{r_{ij}} \right)^6 \right] + \frac{q_i q_j}{4\pi\epsilon_0 r_{ij}}$$

non-bonded

Simplified course of events of MD simulations

- 1) define at t_0
 - all atoms with masses m_i
 - initial atom positions r_i
 - small iteration timestep Δt
 - potential energy U (in the forcefield)
- 2) get accelerations a_i from potential energy U

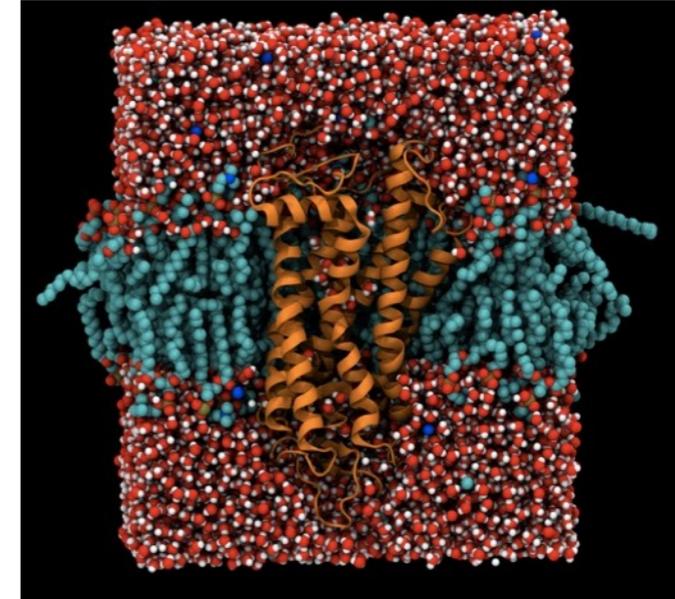
Newton's equations of motion:

$$f_i = m_i a_i \Rightarrow a_i = \frac{f_i}{m_i} ; \quad f_i = -\frac{\partial}{\partial r_i} U ; \quad U = U_{non-bonded} + U_{bonded}$$

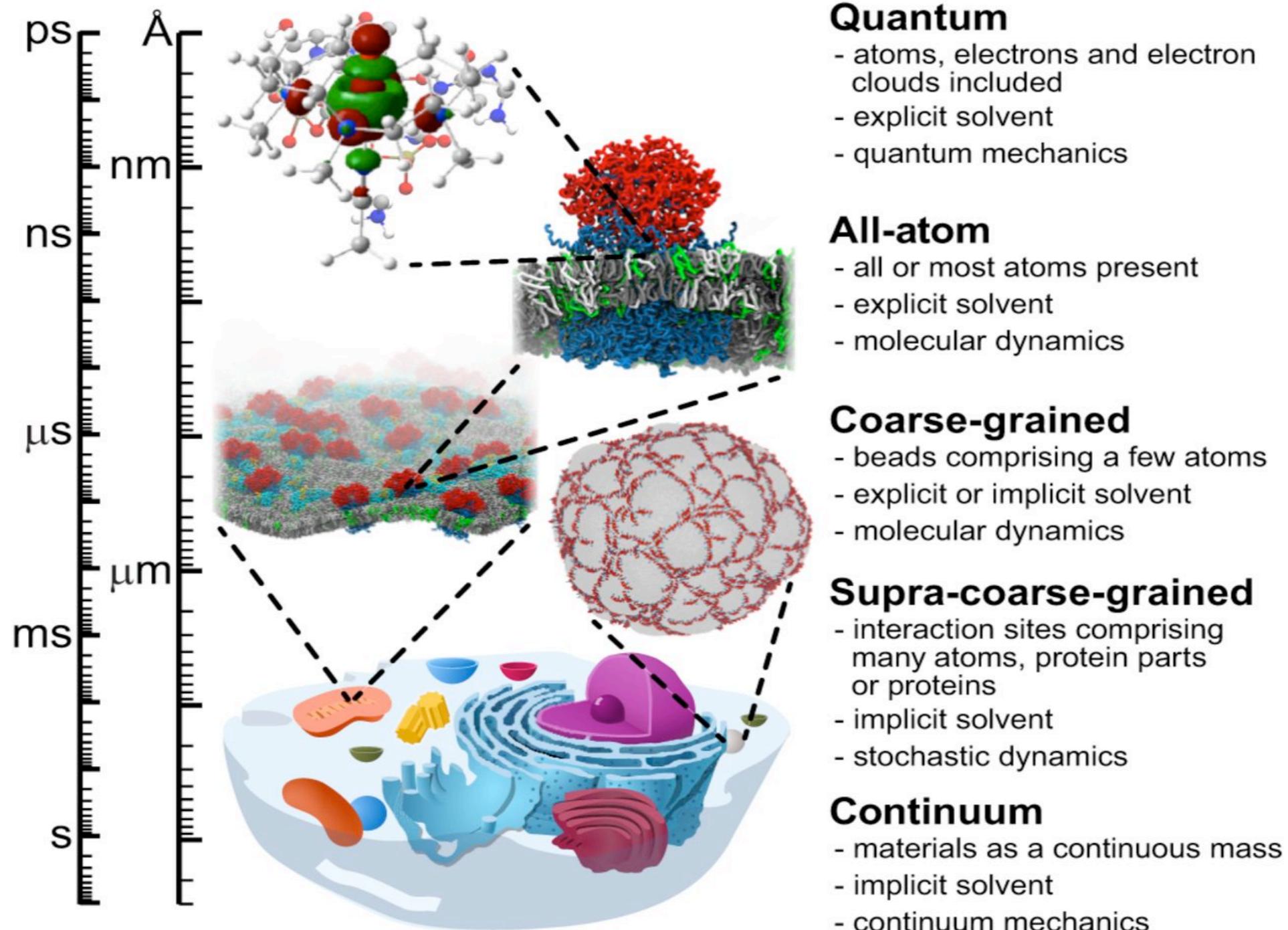
- 3) moving atoms

$$r_{i(t+\Delta t)} = r_i + \frac{1}{2} a_i \Delta t^2 ; \quad a_i = \ddot{r}_i$$

- 4) repeat from 2) till end of simulation



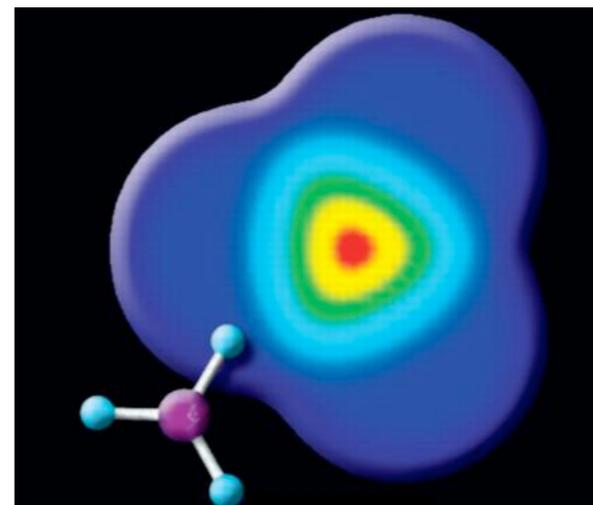
types of MD simulations



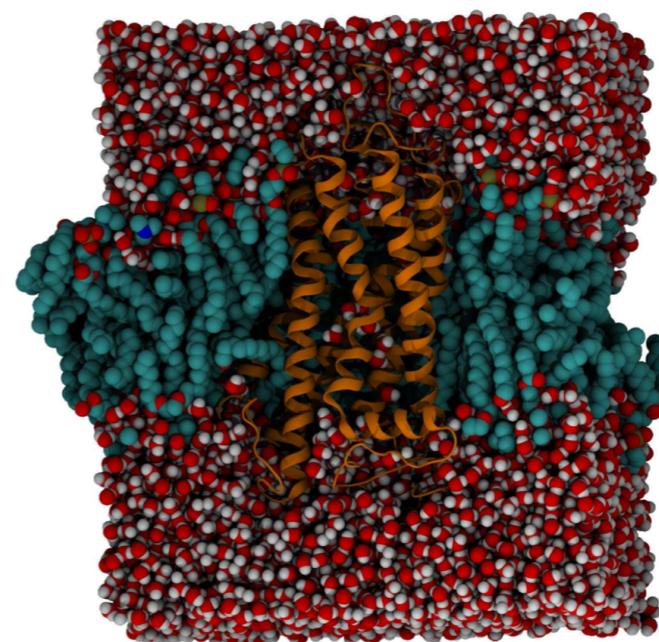
Computational 'microscopy' of cellular membranes Helgi I. Ingólfsson, Clément Arnarez, Xavier Periole, Siewert J. Marrink *Journal of Cell Science* 2016 129: 257-268; doi: 10.1242/jcs.176040

QM or MM?

- **Quantum mechanics (QM):**
- Electronic structure
- slow and computationally expensive
- up to 1000 atoms can be treated
- Chemical reaction can be calculated

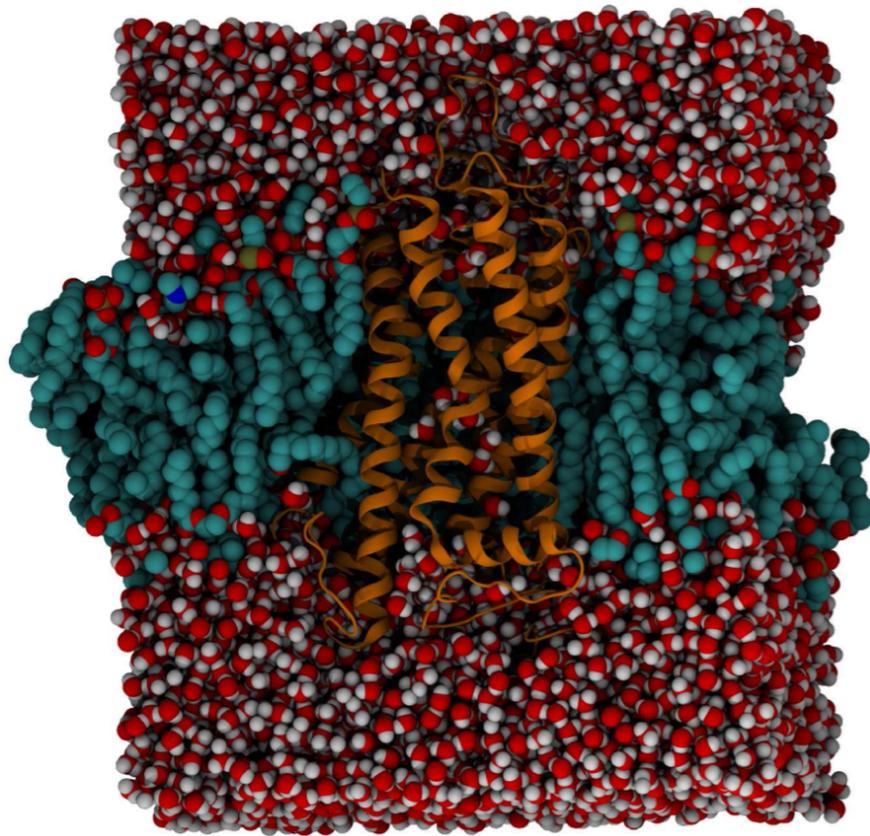


- **Molecular mechanics (MM):**
- Empirical forces, Neglects electronic structure
- computational cheap and fast
- up to 1,000,000 atoms
- Chemical reactions cannot be simulated



Everything Should Be Made as Simple as Possible, But Not Simpler, A. Einstein

MD simulations plus and minus



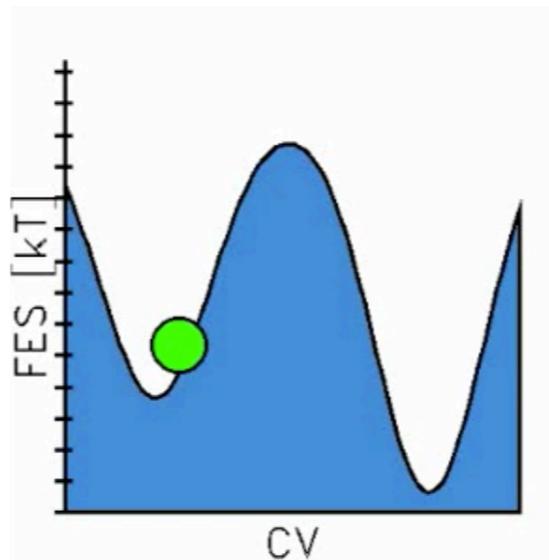
MD simulations overcome several limitations of static computational methods:

- + take explicitly into account solvents and flexibilities
- + apply physical principles + semi-empirical force fields
- + ‚computational microscope‘

- sampling problem: ergodicity is hindered by the form of the system's energy landscape

sampling problem

the system (e.g. protein in simulation box) only samples a local energy minimum and does not reach functionally meaningful energy minima



Movie by Giovanni Bussi

MD simulations: Timescales

„If you do **not observe** a specific structural change
it may always be due to **insufficient sampling**“

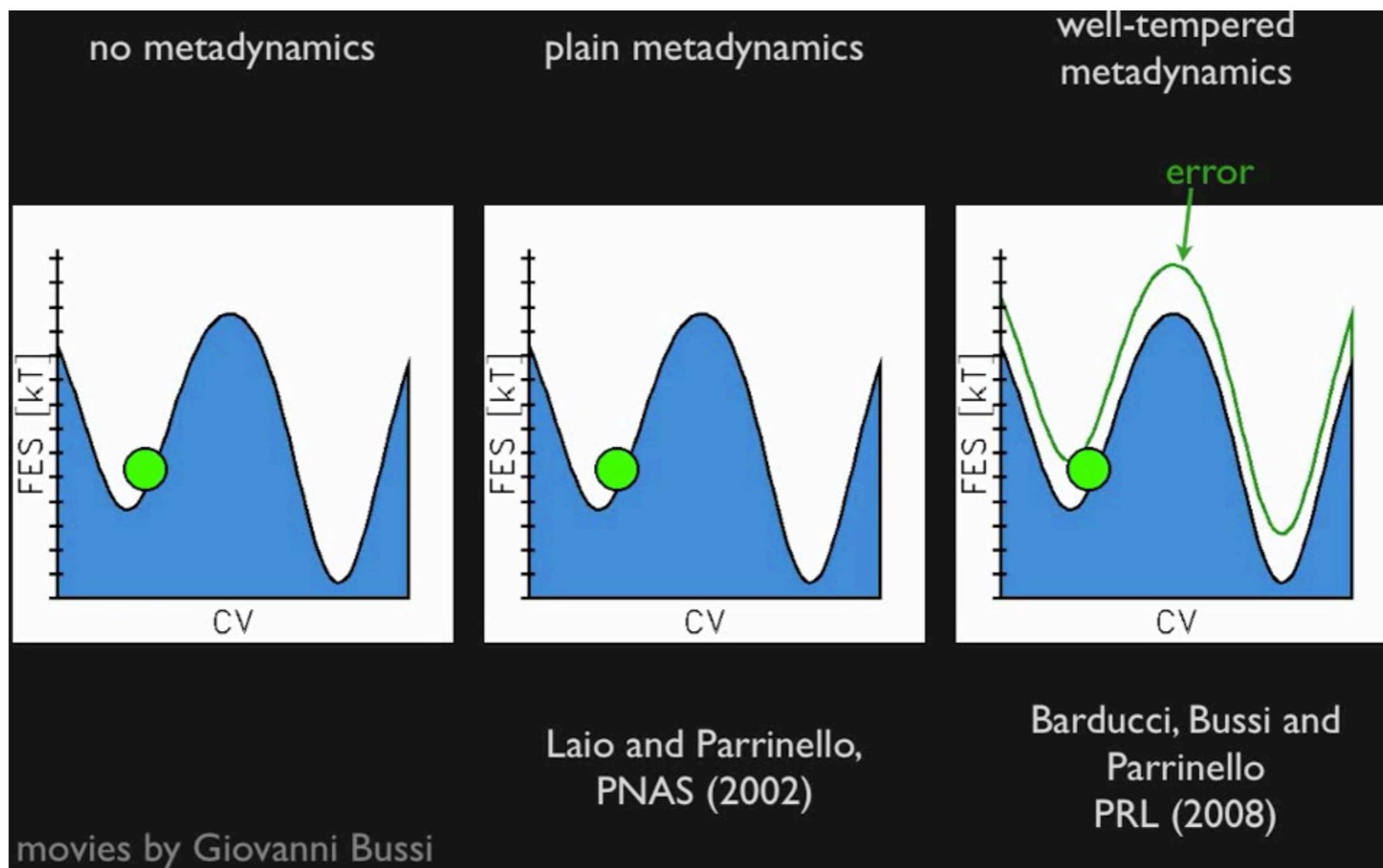
„If you discuss ‚**stability**‘ of a system, always list the limiting
simulation conditions at least **simulation time**“

„If you **observe** a specific structural change only rarely,
always try to **increase sampling**“

MD simulation is stochastic
More than one replica is required

Thermodynamics of ligand binding with metadynamics

- binding free energies along physically meaningful pathways
- thermodynamics and kinetics of binding
- binding path

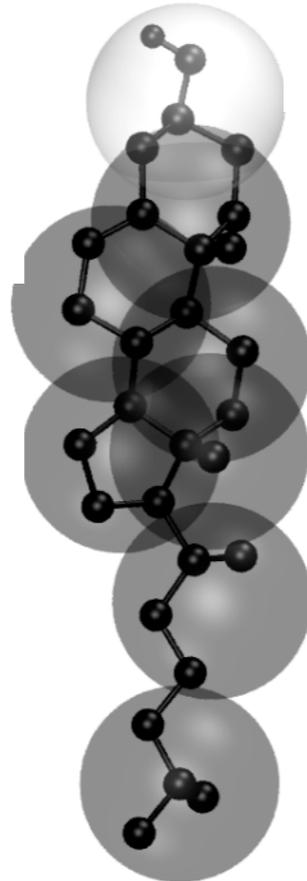


Course Grained Molecular Dynamics Simulations

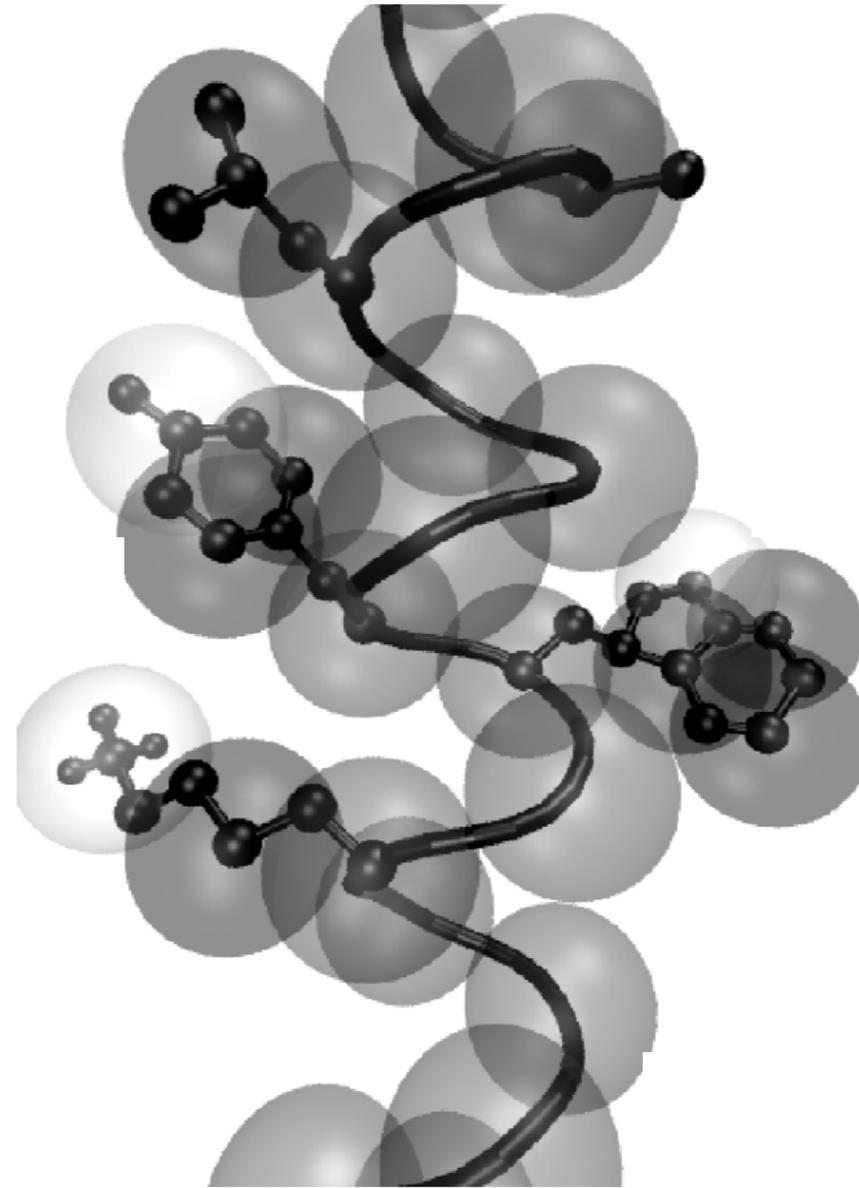
DPPC



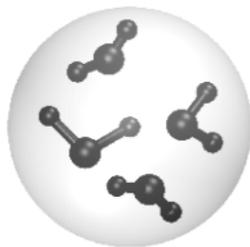
CHOLESTEROL



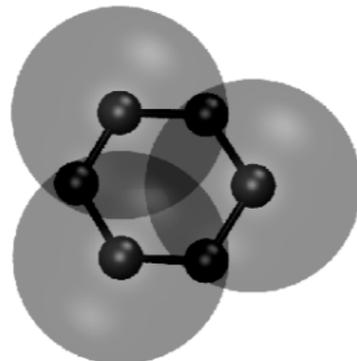
PEPTIDE (ALYWK)



WATER

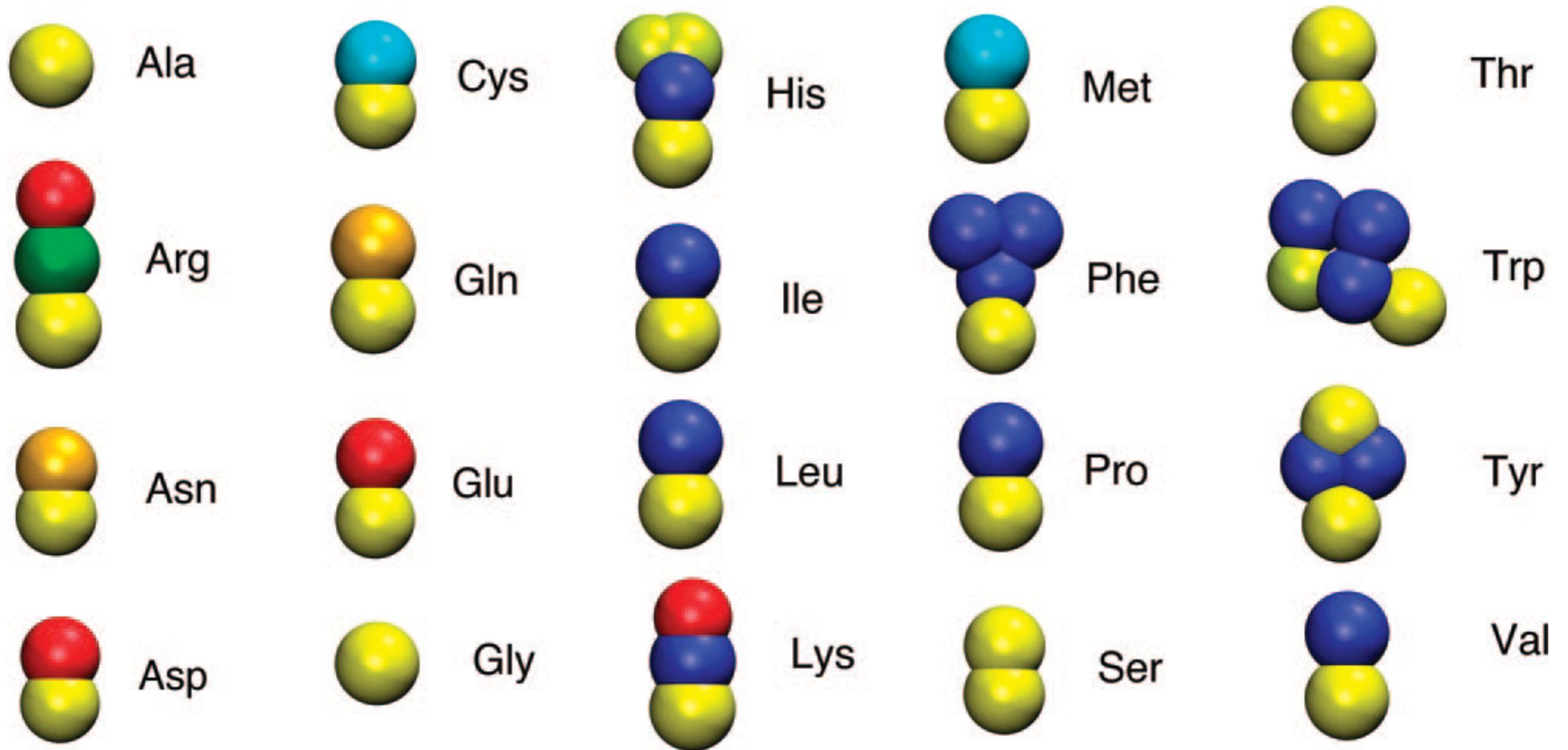


BENZENE



Martini Homepage (Marrink), <http://md.chem.rug.nl/cgmartini/index.php/about>

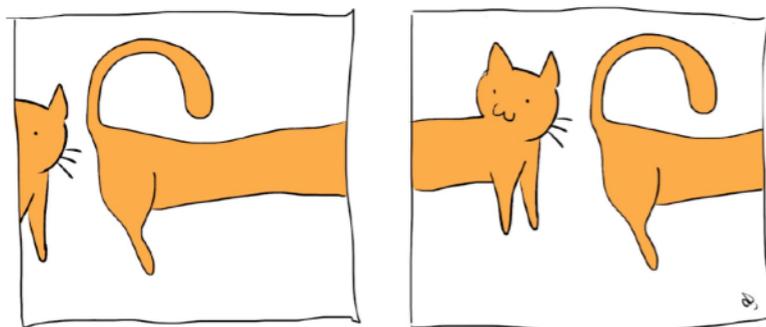
Course Grained Molecular Dynamics (CGMD) Simulations



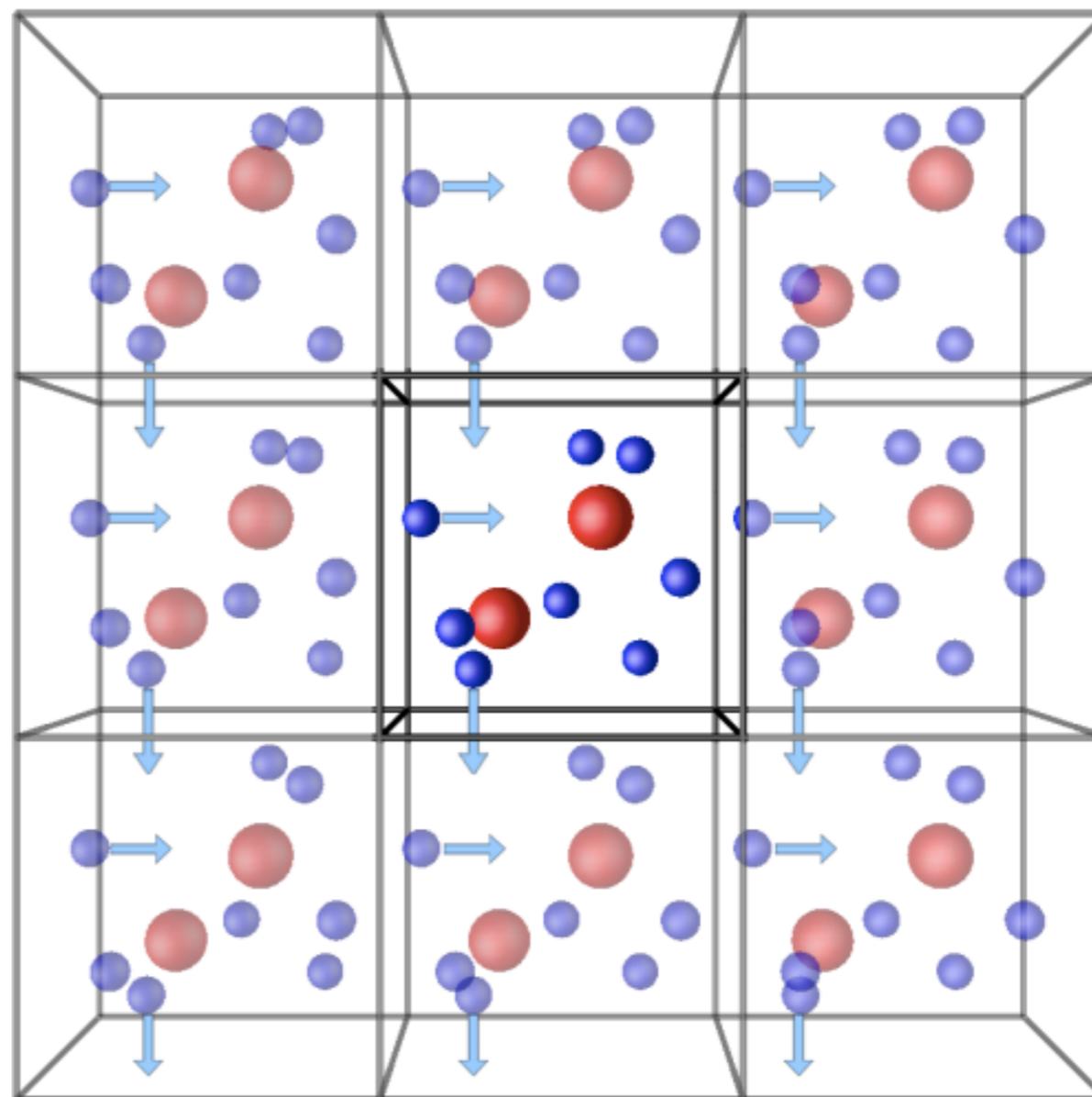
Monticelli et al., 2008, J. Chem. Theory and Comput.

Periodic boundary conditions (schematic)

PERIODIC BOUNDARY CONDITIONS



a particle leaving the simulation
box at one side
-> enters the box at the opposite
side
(in the 3 dimensional space)



<http://isaacs.sourceforge.net/phys/psc.html>

Periodic boundary conditions (cinematic)



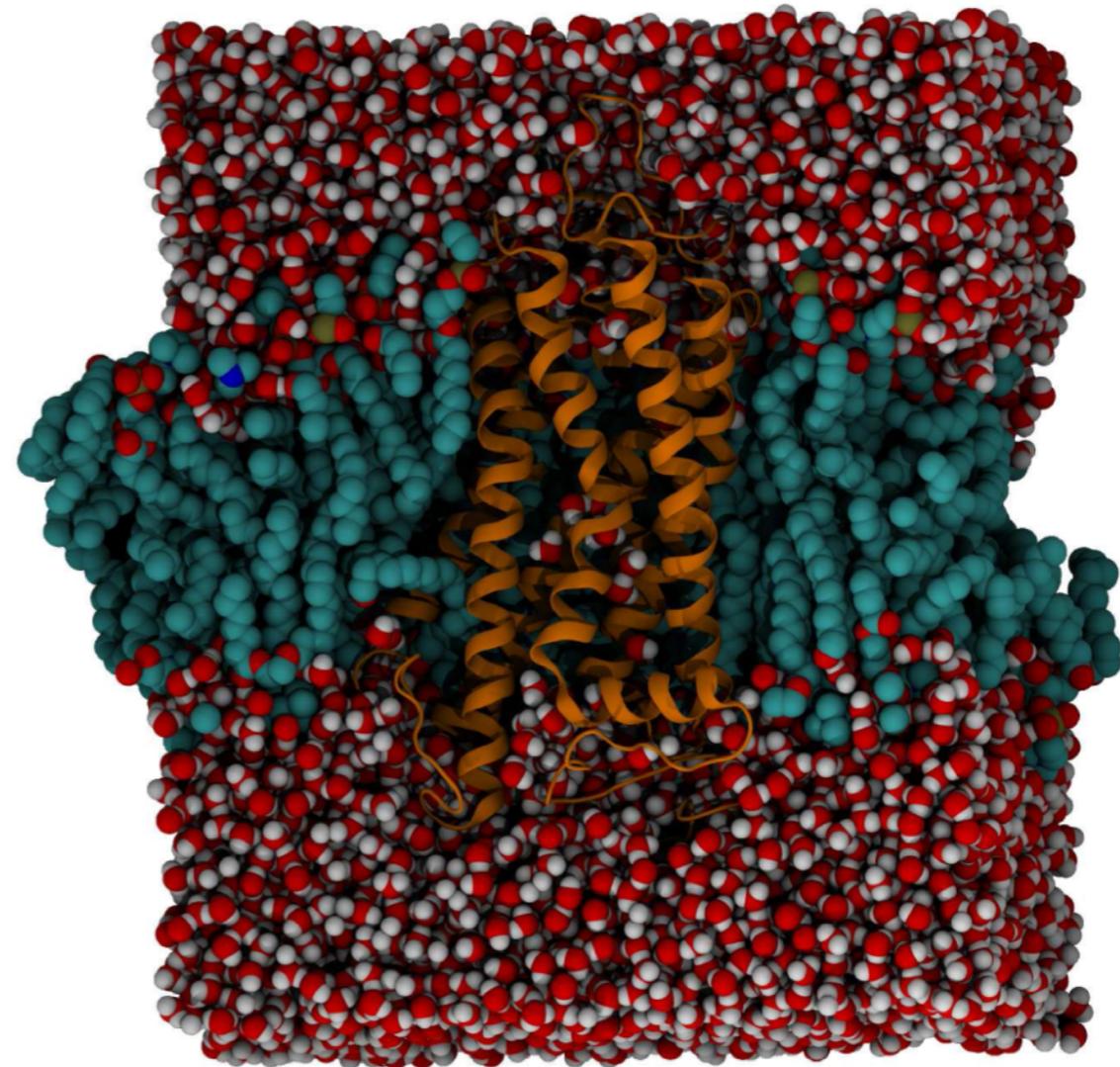
<http://isaacs.sourceforge.net/phys/pbc.html>

approximating the behavior of macro-scale system

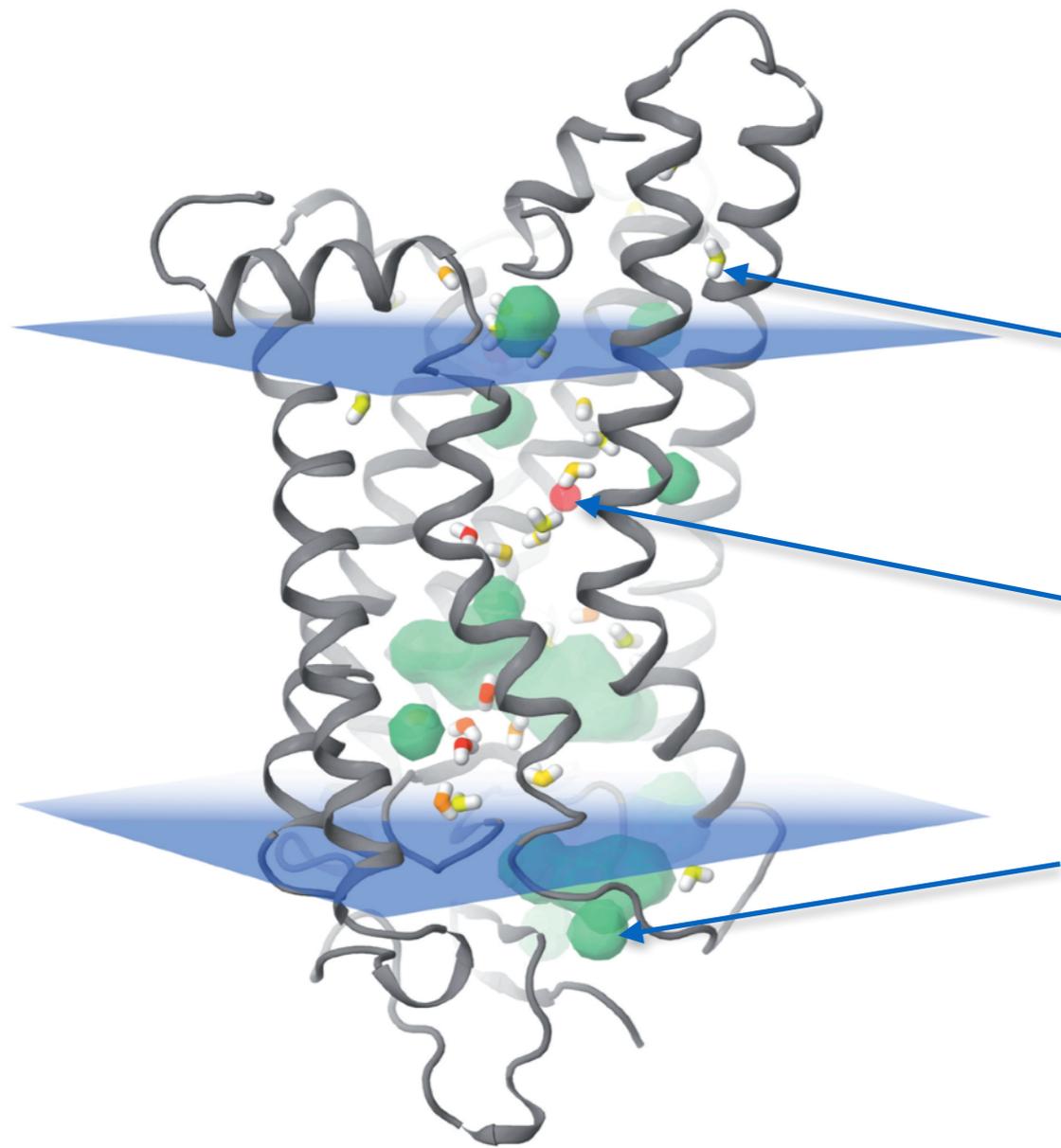
Box size depends on the size of the system e.g. here the solvated receptor embedded in a lipid bilayer

The net electrostatic charge of the system must be zero:

- add Na^+ or Cl^- to approximate the ionic strength of the solution



Setup MD simulations: Internal waters / cavities



sticks colored from red (30 kcal/mol) to yellow (10 kcal/mol)

water determined by crystal structure depicted as translucent red ball

,hydrophobic cavities depicted as translucent green surfaces

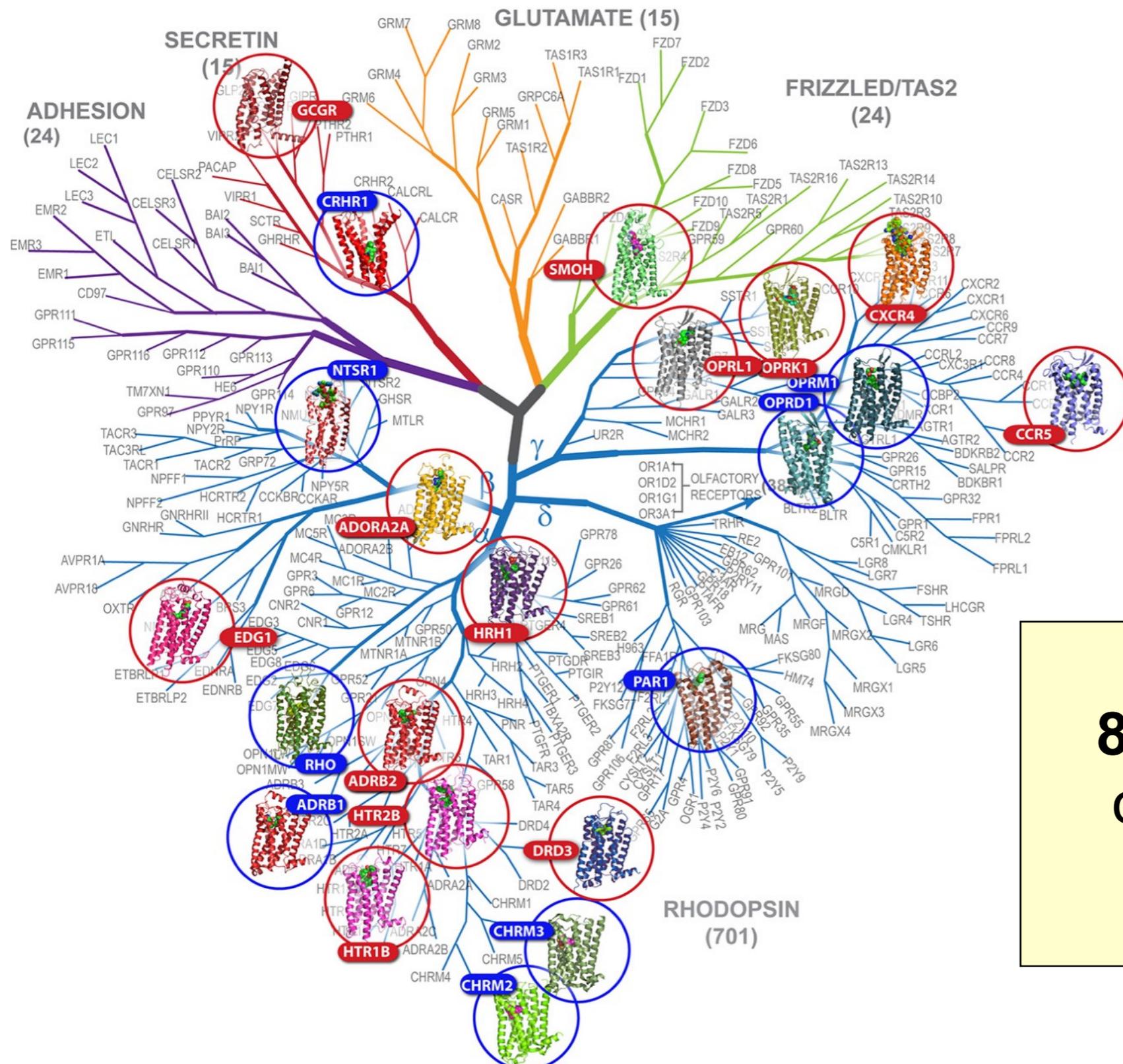
<http://proteinformatics.uni-leipzig.de/mppd/>

Rose, Theune, Goede & Hildebrand, *NAR* 2009

<http://lmc.uab.es/homolwat>

Mayol et al, *NAR* 2020

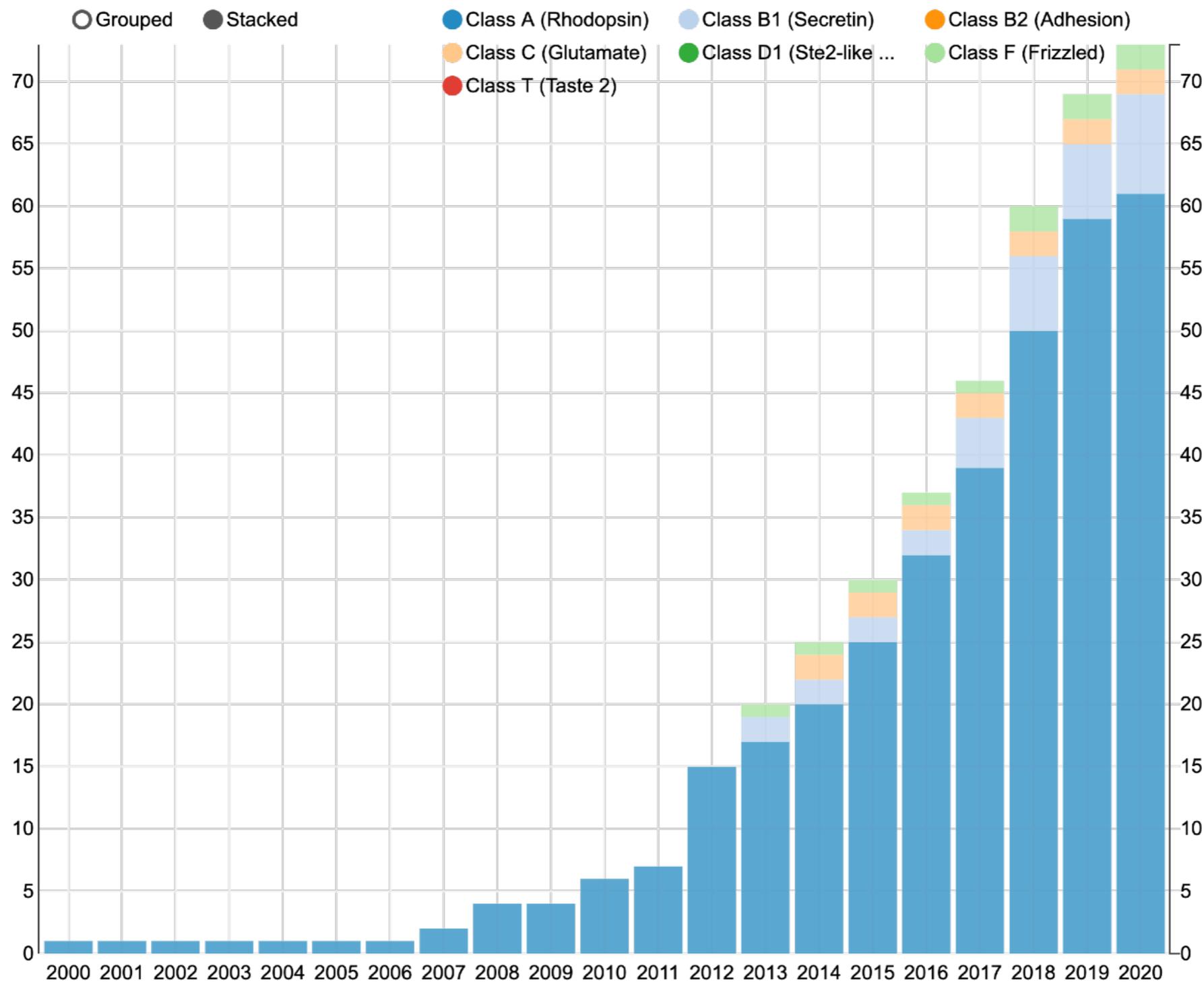
...many (crystal) structures resolved



850 receptors but only 73 resolved

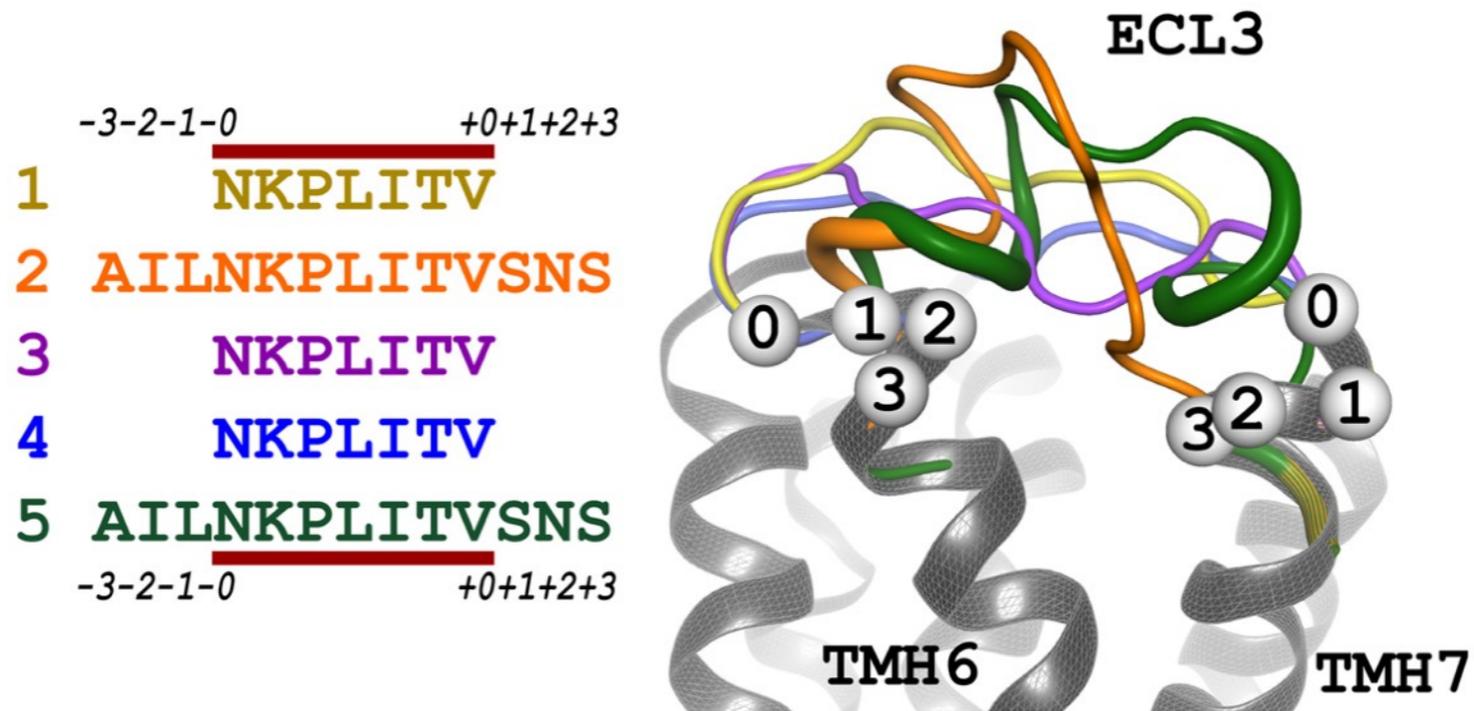
GPCR Network: <http://gpcr.scripps.edu>

Titeltext ...many (crystal) structures resolved



GPCR DB: <https://gpcrdb.org/structure/statistics>

Loopmodelling

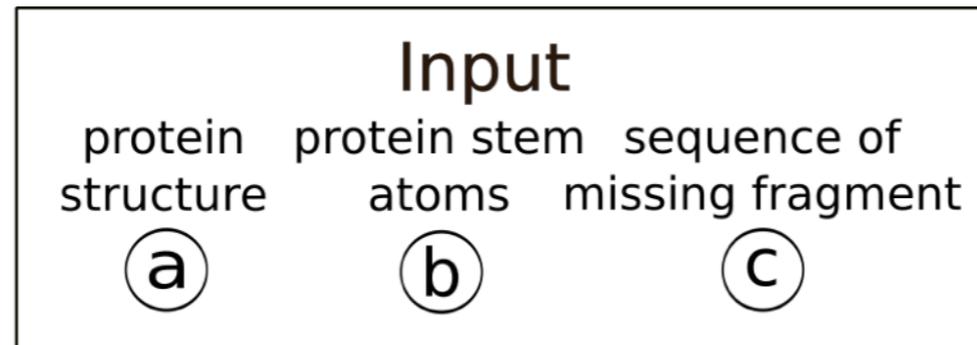


#	Ext	GPCR	Score	Seq. Ident.	PDB ID
1	0	✓	0.096	28.57	5C1M
2	3	✓	0.095	30.77	3PBL
3	0	✓	0.083	28.57	4K5Y
4	0	✓	0.093	28.57	4XT1
5	3	✓	0.068	15.38	4Z43

<http://www.ssfa-7tmr.de/ssfe2/>

Worth et al., *NAR* 2009

SuperLooper, An Interactive webtool



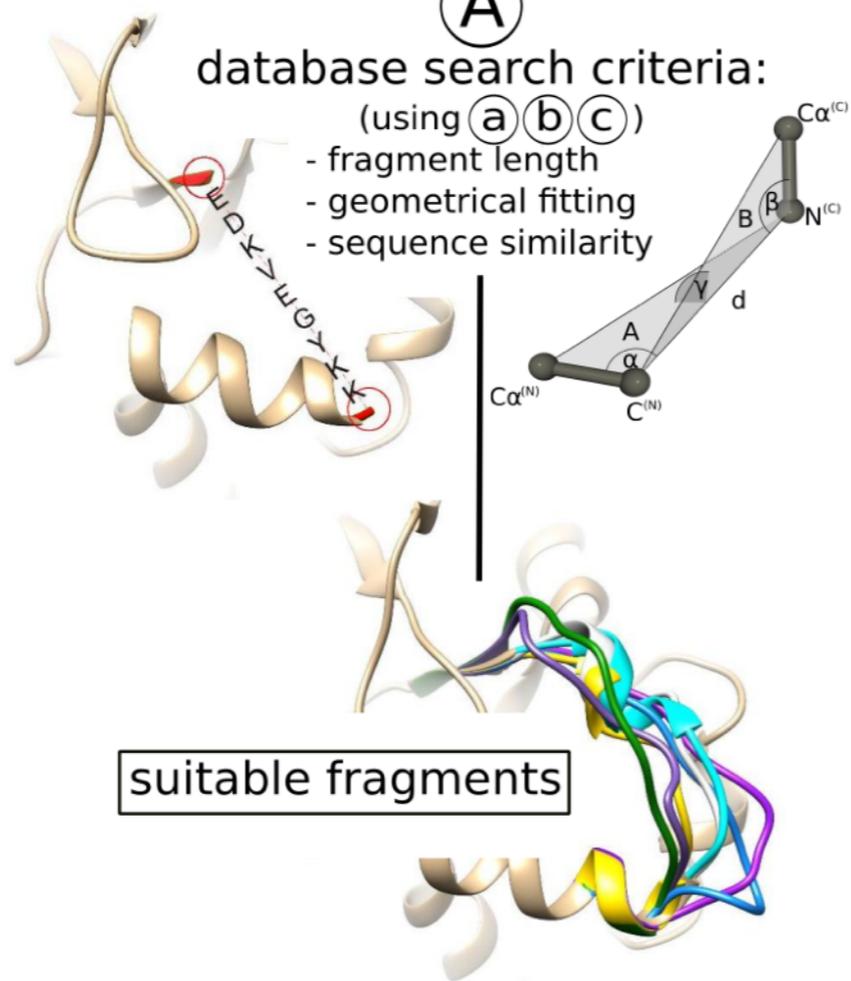
FragSearch

(A)

database search criteria:

(using (a) (b) (c))

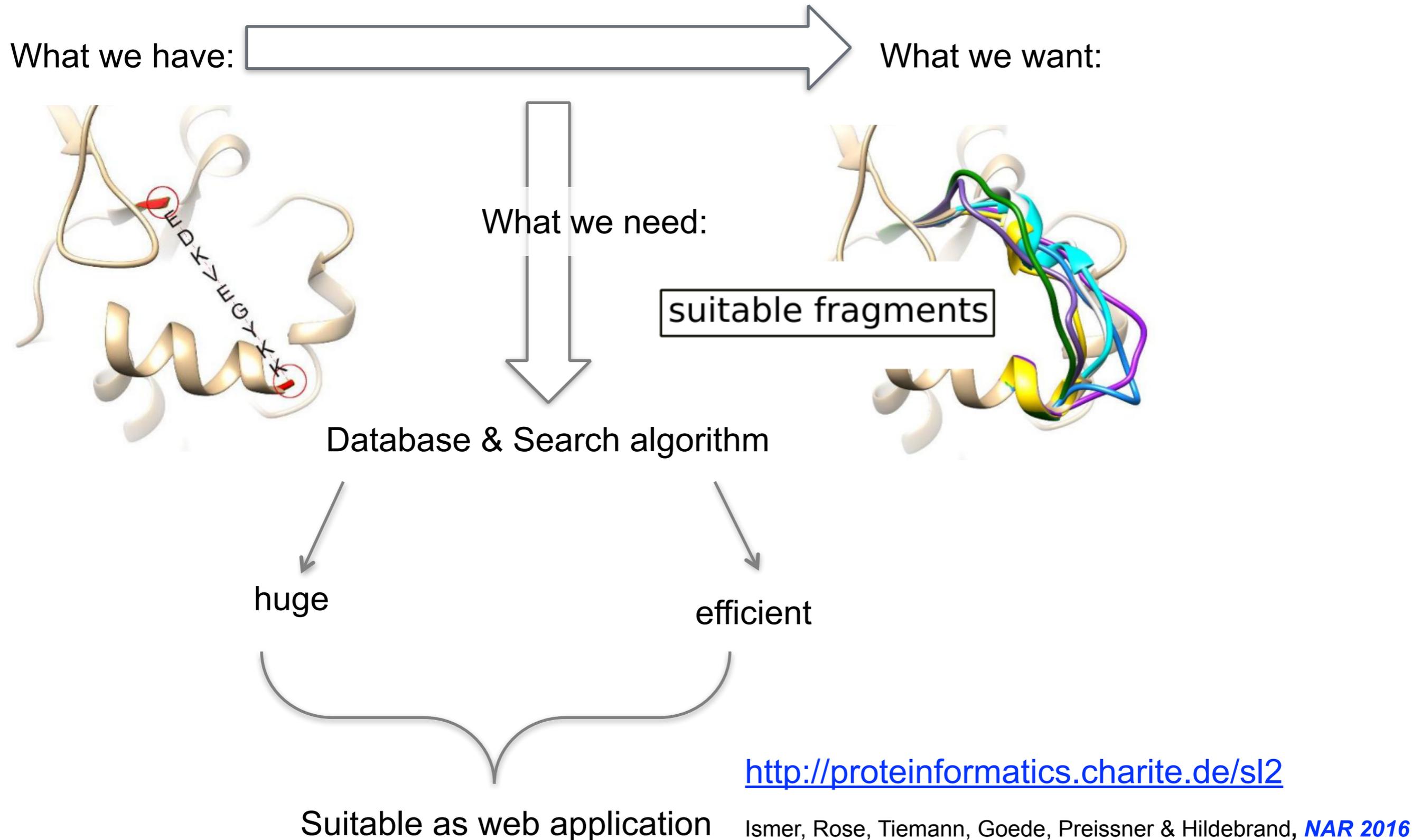
- fragment length
- geometrical fitting
- sequence similarity



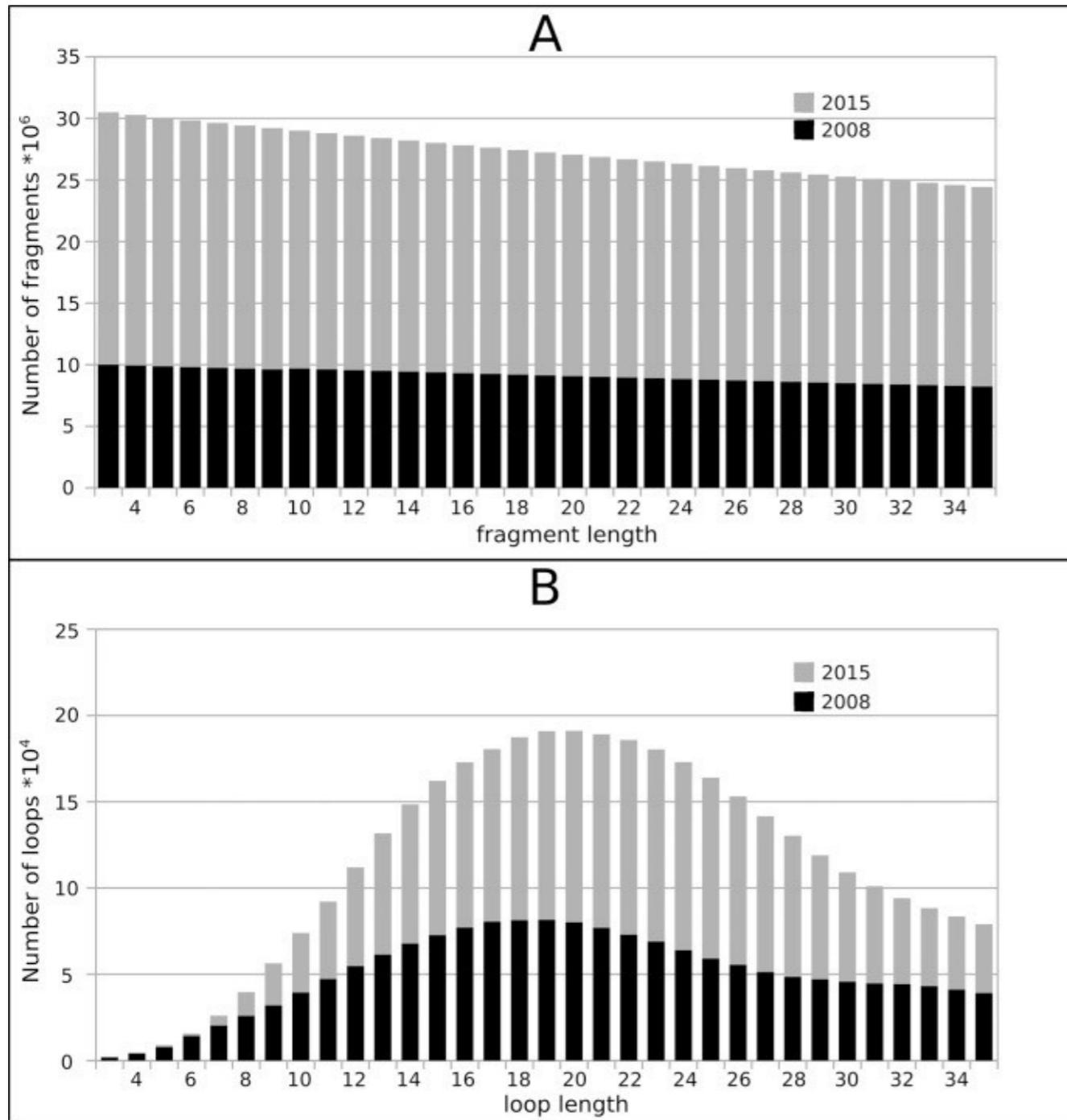
<http://proteinformatics.charite.de/sl2>

Ismer, Rose, Tiemann, Goede, Preissner & Hildebrand, *NAR* 2016

SuperLooper, An Interactive webtool



SuperLooper, An Interactive webtool



SL2 – Database

- 114,693 structures
- 901,609,231 fragments
- 3-35 residue length

<http://proteinformatics.charite.de/sl2>

Ismer, Rose, Tiemann, Goede, Preissner & Hildebrand, *NAR* 2016

SuperLooper, An Interactive webtool

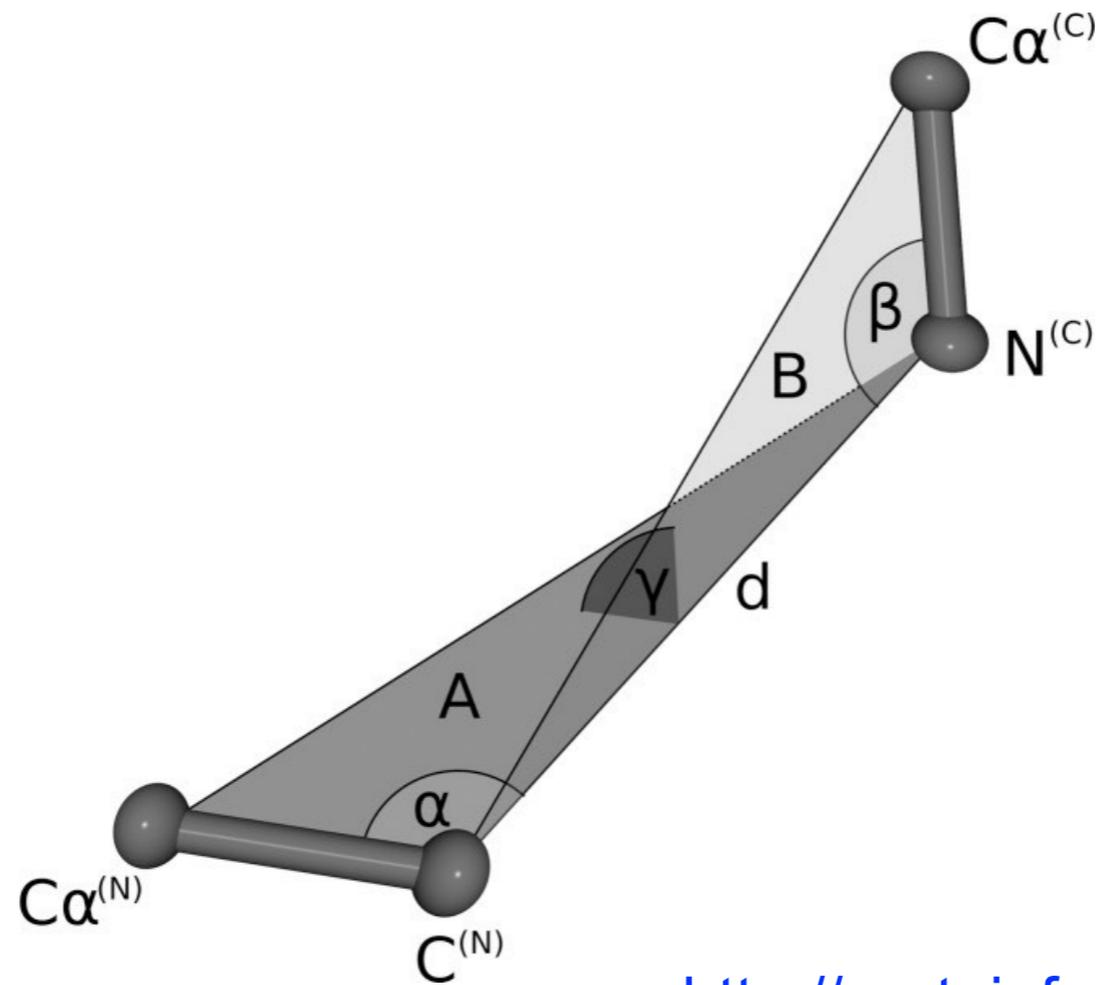
SL2 – Search algorithm / criteria

1. Extract fragments with sequence length of missing fragment
2. Weight fragments according to sequence similarity
3. Rank fragments according to geometrical fingerprint matching (RMSD > 0.75 Å)
4. Sort similar fragments with backbone RMSD < 0.5 Å of top-1000 list out
5. Find clashes
6. Present results to user

<http://proteinformatics.charite.de/sl2>

Ismer, Rose, Tiemann, Goede, Preissner & Hildebrand, *NAR* 2016

Geometrical fingerprint



<http://proteininformatics.charite.de/sl2>

Ismer, Rose, Tiemann, Goede, Preissner & Hildebrand, *NAR* 2016

SuperLooper, An Interactive webtool

The screenshot displays the SuperLooper webtool interface. On the left, a 3D ribbon diagram of a protein structure is shown, with a central stem highlighted in green and a horizontal membrane plane in brown. The protein is rendered in grey. The interface includes a menu bar with 'View' and 'Help', and a title bar 'NGL Viewer 0.7dev'. The main control panel on the right contains the following elements:

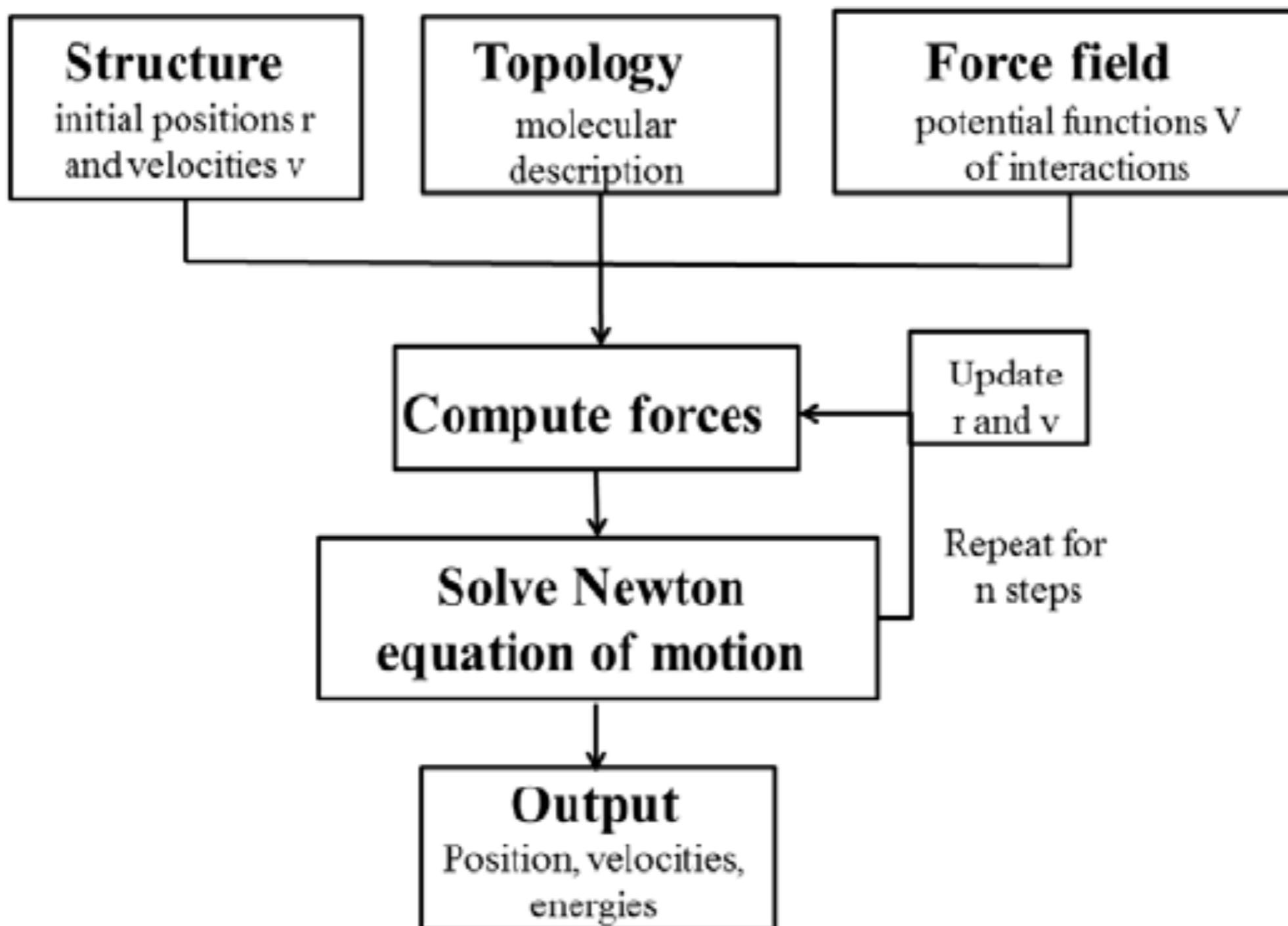
- Buttons: 'New SL2 Job', 'Load SL2 Example', 'Submit', 'Stop', 'Calculate membrane plane', 'Clear input fields', 'Decrease N-terminal stem', 'Increase C-terminal stem'.
- Form fields: 'PDB File' (with 'Browse...' and '4xt1.pdb'), 'Stem residue 1' (with '32:B'), 'Stem residue 2' (with '103:A'), 'Loop sequence' (with 'DHNSLASV'), and 'MembraneDB' (checkbox).
- Color scheme: 'Score' (dropdown menu).
- Buttons: 'show all', 'show first', 'download all'.
- Table with columns: ID, Dc, Sc..., PDB, Cl..., Sequence, Se..., Seq Position.

ID	Dc	Sc...	PDB	Cl...	Sequence	Se...	Seq Position
1	↓	0.16	1s7l	0		50.00	16-23:A
2	↓	0.16	2h7g	1	DKNFLDVV	50.00	285-292:X
3	↓	0.15	4abt	15	DGNILATN	50.00	18-25:A
4	↓	0.15	3a9v	1	DKHDLSSL	37.50	292-299:A
5	↓	0.14	1u1v	2	DHRALSNF	37.50	172-179:A
6	↓	0.14	2r3v	2	DVARLQSL	37.50	63-70:A
7	↓	0.13	5c5i	1	DPADLAEL	37.50	54-61:A
8	↓	0.12	4ff6	0	DNVDLSDI	25.00	63-70:A
9	↓	0.12	4j3s	0	DECELATF	37.50	326-333:A
10	↓	0.12	2pod	2	SLSSLKRY	25.00	251-258:A
11	↓	0.11	2w8d	1	DSSDVTEV	25.00	216-223:B
12	↓	0.11	1wvl	2	SPDELPQV	25.00	293-300:B
13	↓	0.11	2bt2	0	SFDLLSS	25.00	65-72:E
14	↓	0.11	1r9j	1	DgSTLSLF	25.00	190-197:B
15	↓	0.11	3oi8	1	EQFHLKSI	25.00	121-128:A
16	↓	0.11	3u94	2	SADDLAKQ	25.00	122-129:A
17	↓	0.11	1p59	0	DRDELMKL	25.00	108-115:A
18	↓	0.11	3rle	9	ggQgLLgV	25.00	91-98:A

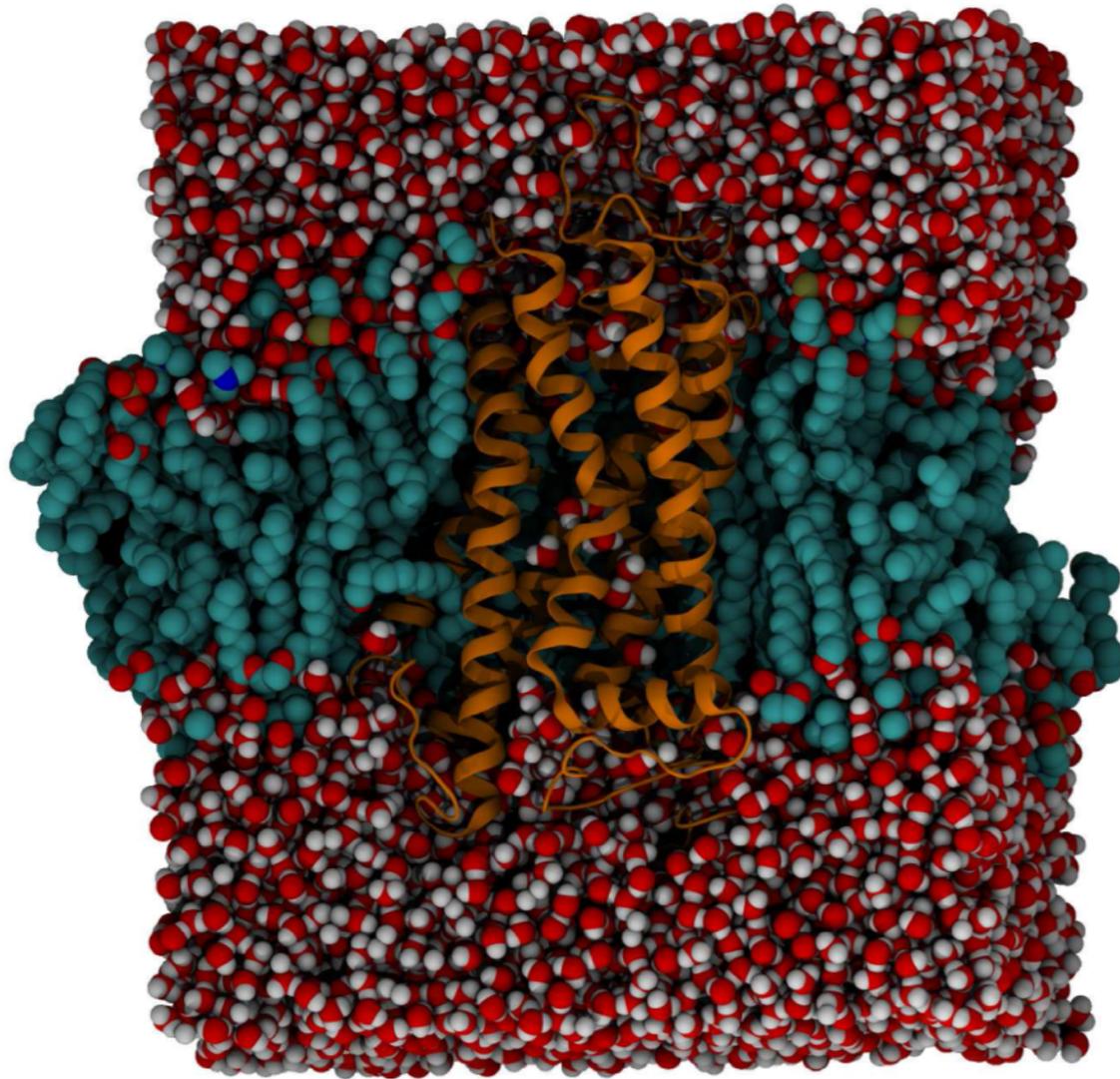
<http://proteininformatics.charite.de/sl2>

Ismer, Rose, Tiemann, Goede, Preissner & Hildebrand, *NAR* 2016

Simplified course of events of classical MD simulations



Setup MD simulations: setup



- MD simulations with GROMACS
- AMBER99SB forcefield

- Loop modelling with SuperLooper

www.proteinformatics.charite.de/SL2

NAR 2009, 2016, 2018

- Intra-molecular waters

www.proteinformatics.charite.de/MPPD

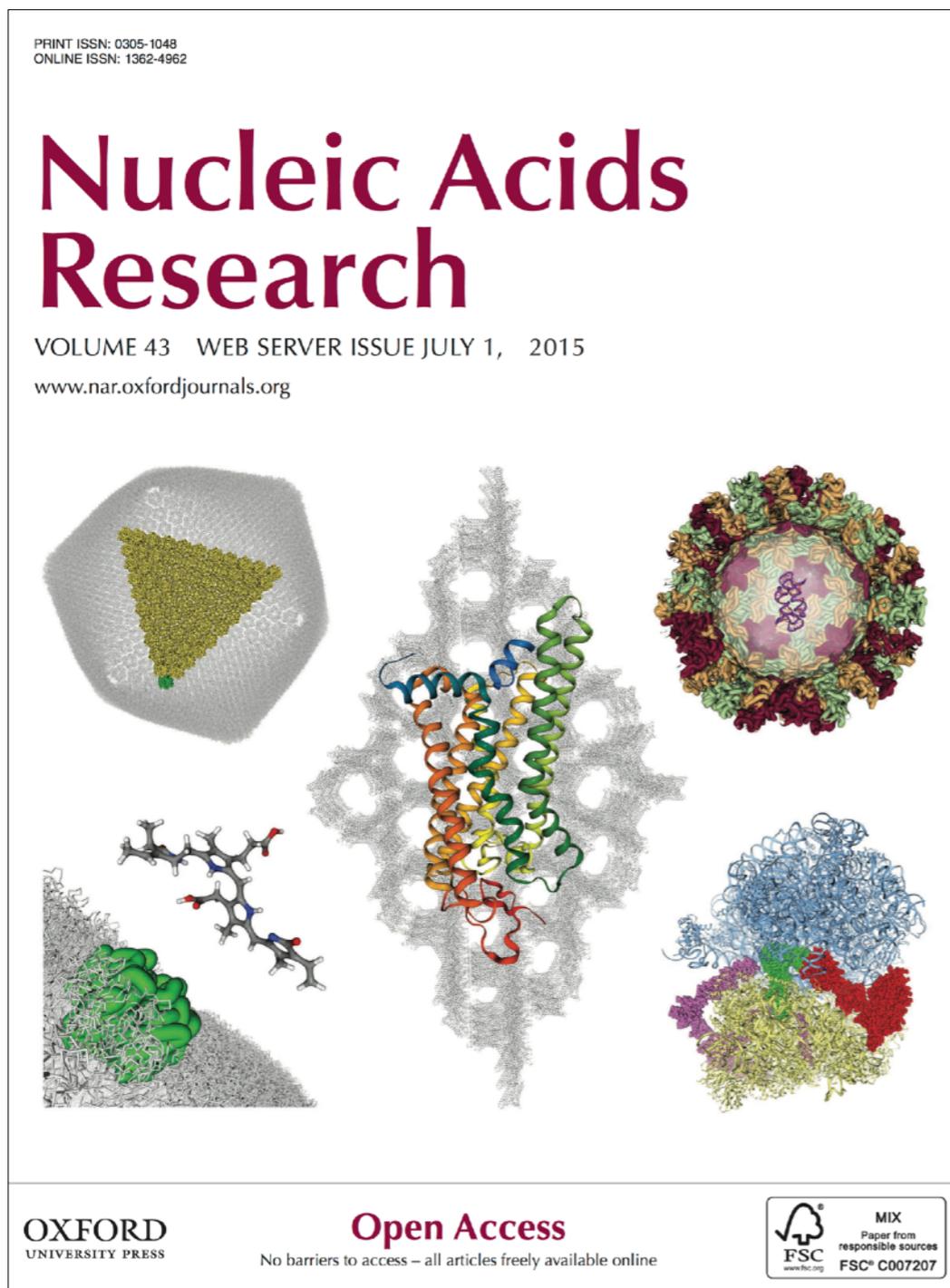
NAR 2014; 2020

- Visualization of trajectories in the WEB

www.proteinformatics.charite.de/MDsrv

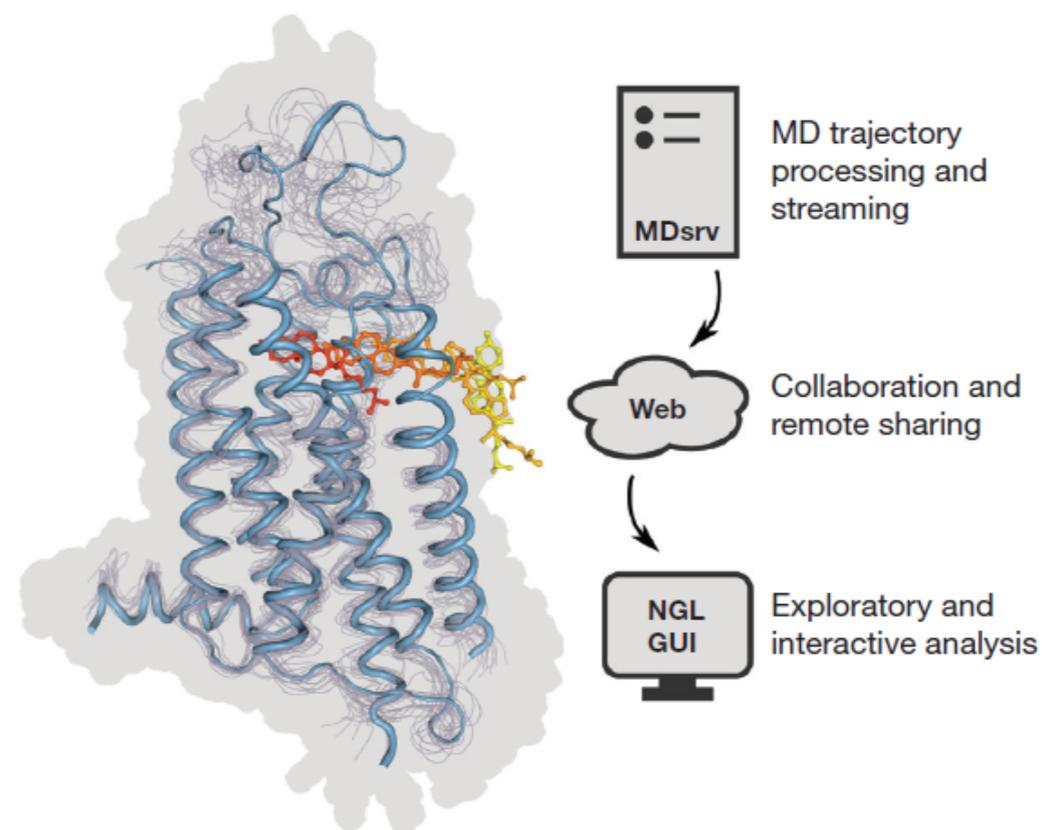
Nature Methods 2017; TIBS 2019

Sommer, Elgeti, **Hildebrand**, Szczepek, Hofmann & Scheerer, *Methods Enzymol* 2015



MDsrv: viewing and sharing molecular dynamics simulations on the web

CORRESPONDENCE

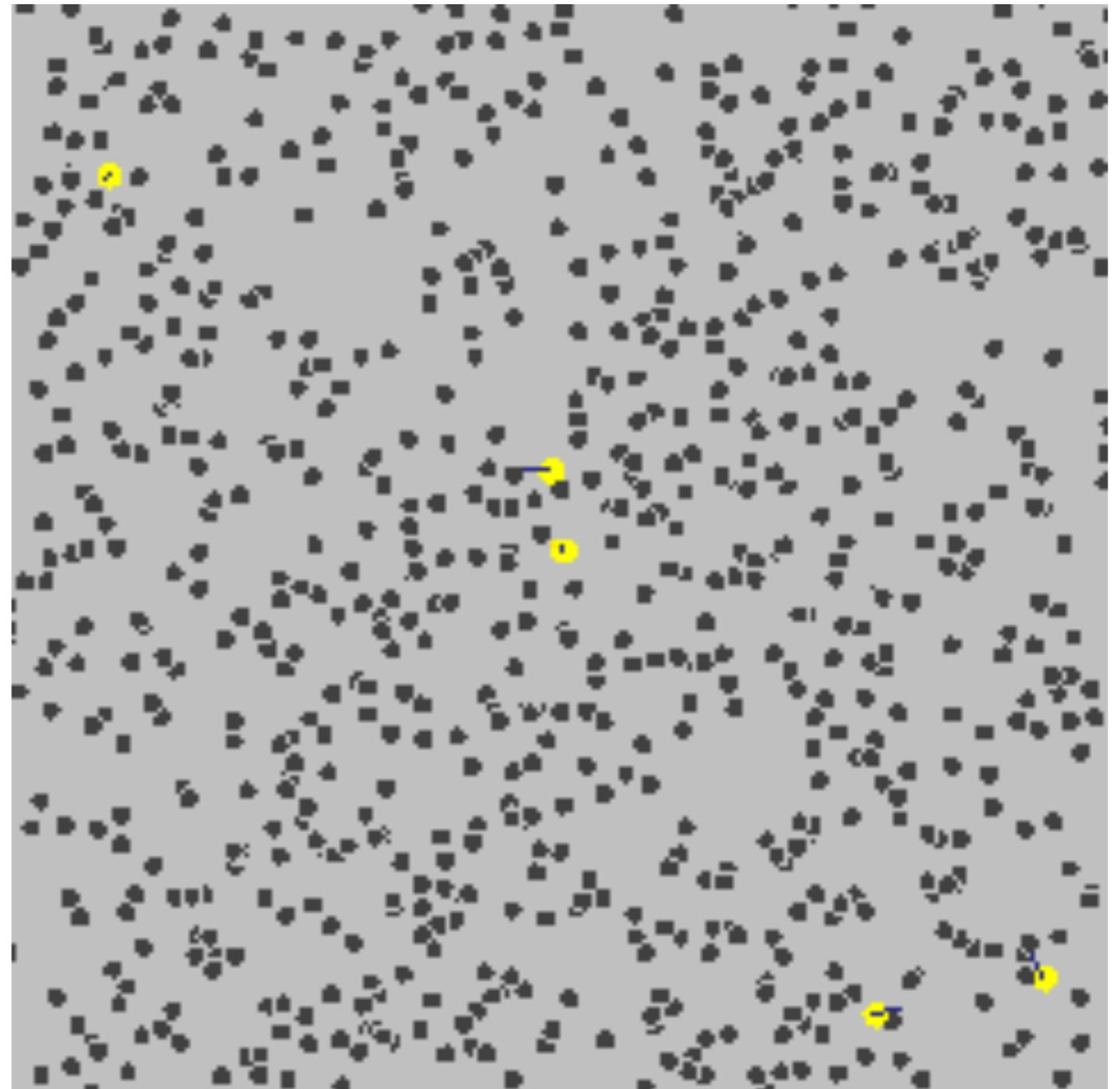


Thermodynamic temperature

Brownian Motion

Thermal motion of particles
in liquids or Gases

By *botanics*
Robert Brown
1827



<https://commons.wikimedia.org/w/index.php?curid=19140415>

Temperature correlates with kinetic energy

$$\frac{m}{2} \bar{v}^2 = \frac{3}{2} k T$$

T - absolute Temperature

refers to an absolute zero according to the properties of the **ideal gas**

k_B - Boltzmann-constant

k = 1,38 · 10⁻²³ J/K

m = Masse

S = k_B ln W Boltzmann-formula

entropy **S** and the number of ways the atoms or molecules of a thermodynamic system arrange

Thermodynamics: Dance of the proteins



$$\Delta G = \Delta H - T^* \Delta S$$

Gibbs-Helmholtz
Equation

Derived from ideal gases

$$p V = n R T$$

Pressure p

Temperatur T

Volumen V

n - number of particles

R - Gas constant

$$R = 8,314 \frac{\text{J}}{\text{mol K}}$$

$$\frac{p_1 V_1}{T_1} = \frac{p_2 V_2}{T_2}$$

Thermodynamics

$$pV = nRT$$

Isothermic $T = \text{const}$

Isobaric $p = \text{const}$

Isochoric $V = \text{const}$

$$p \cdot V = \text{const}$$

$$V \sim T$$

$$p \sim T$$

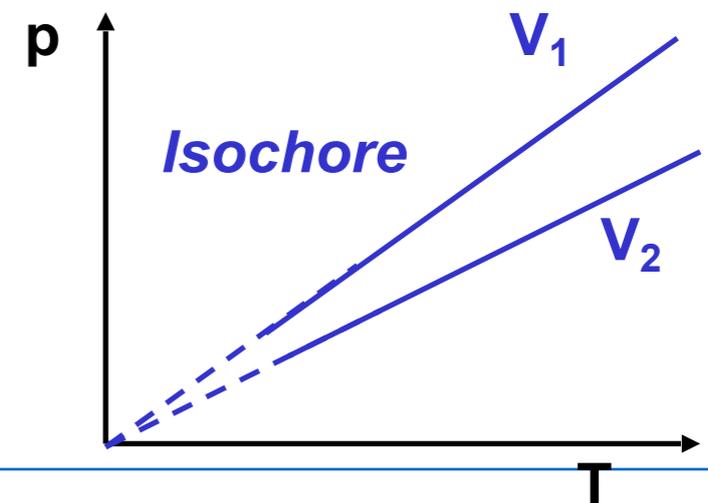
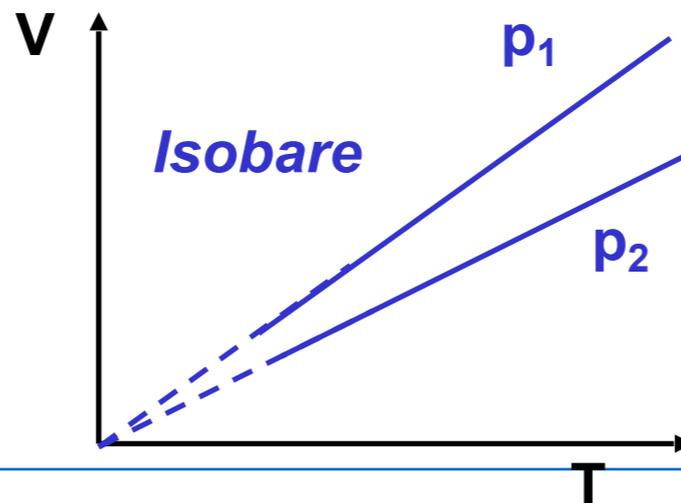
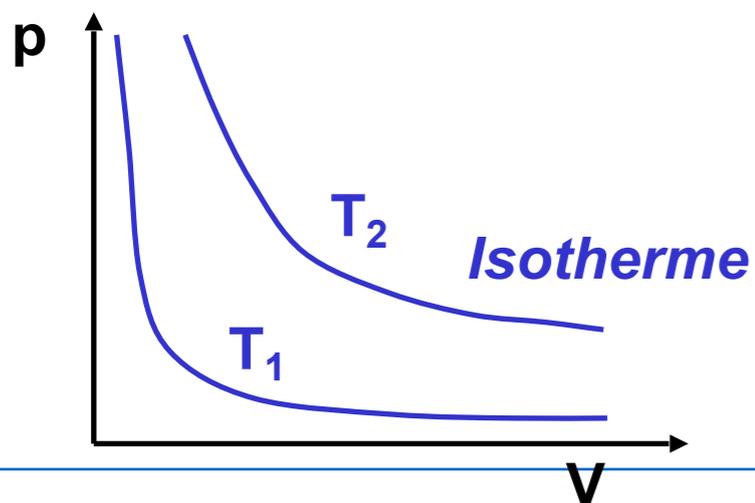
Boyle-Mariotte

Gay-Lussac

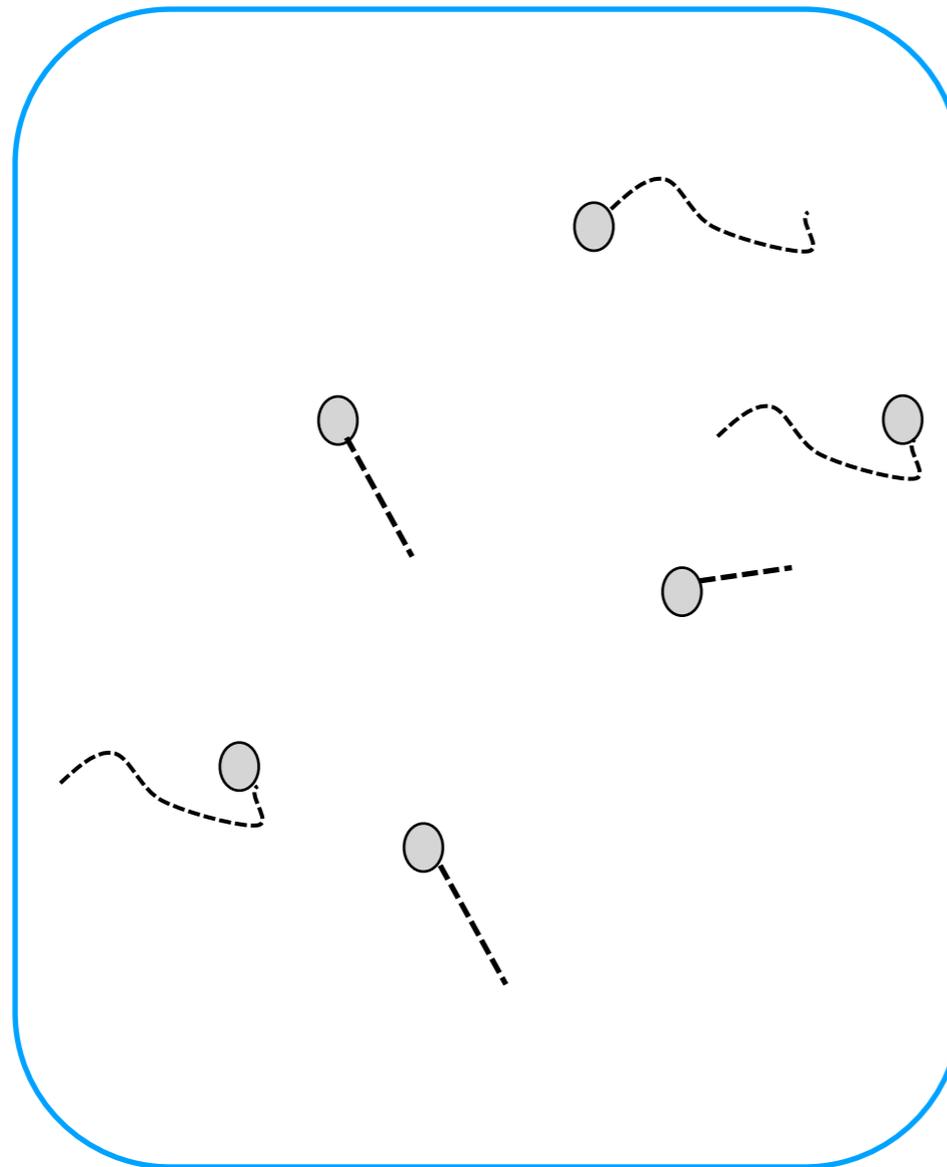
Compression of gas

Extension of gas

Warming up gas in a closed box

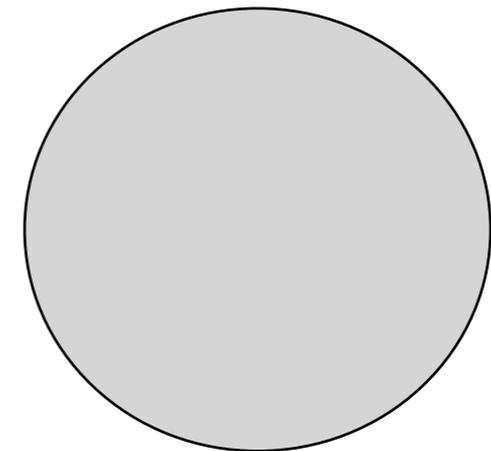


Microscopic toward macroscopic



Microscopic world

Position, velocity, mass



Macroscopic world

Pressure, Volume, Temperature, moles

Molecular Dynamics Ensembles

Constant number of Particle, constant Volume, constant Energy (NVE)

Constant number of Particle, constant Volume, constant Temperature (NVT)

Constant number of Particle, constant Pressure, constant Temperature (NPT)

Choose the ensemble that best fits for the system you want to simulate

The Ergodic hypothesis states

Experiment

MD Simulation

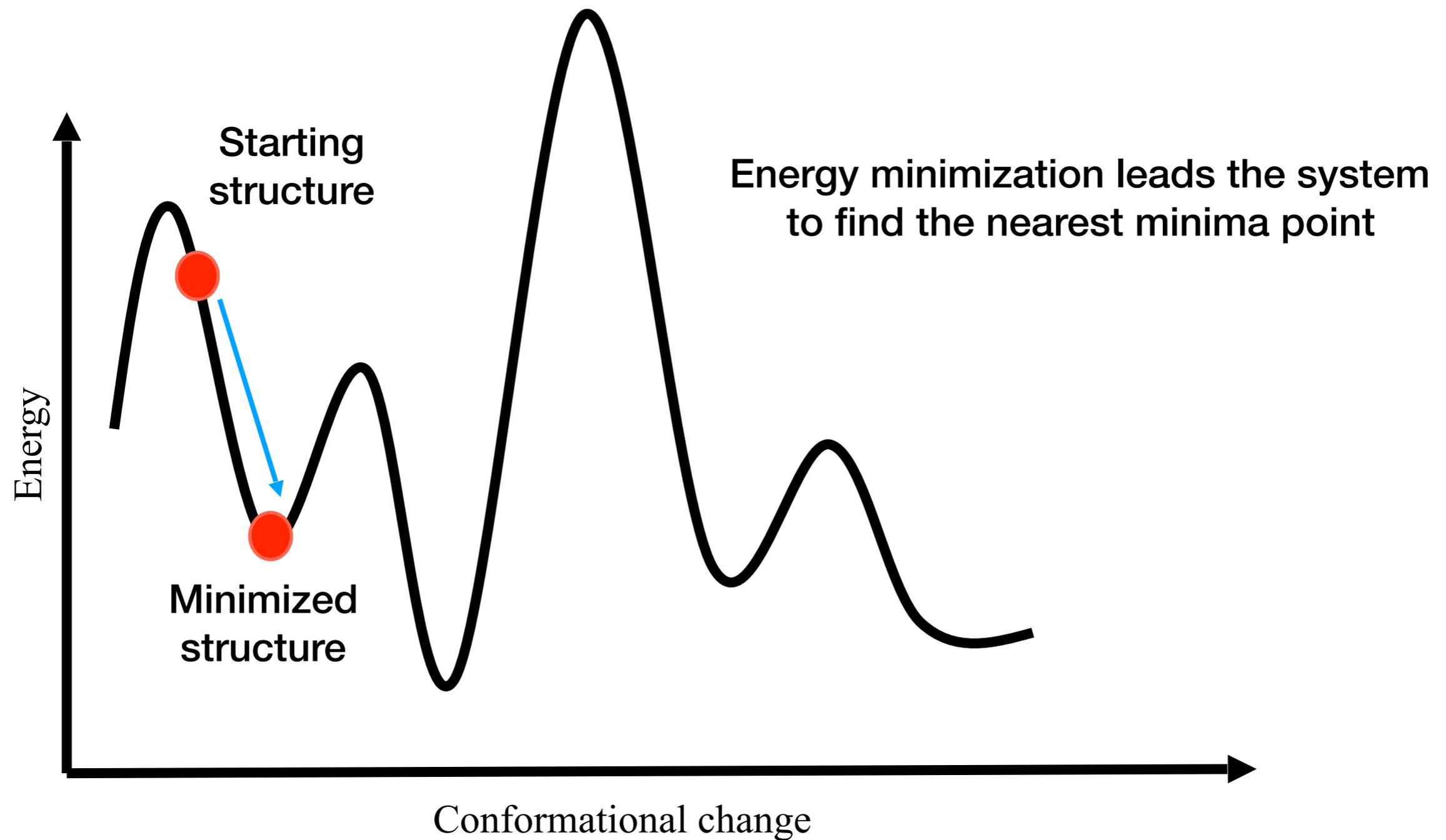
Macroscopic

Microscopic

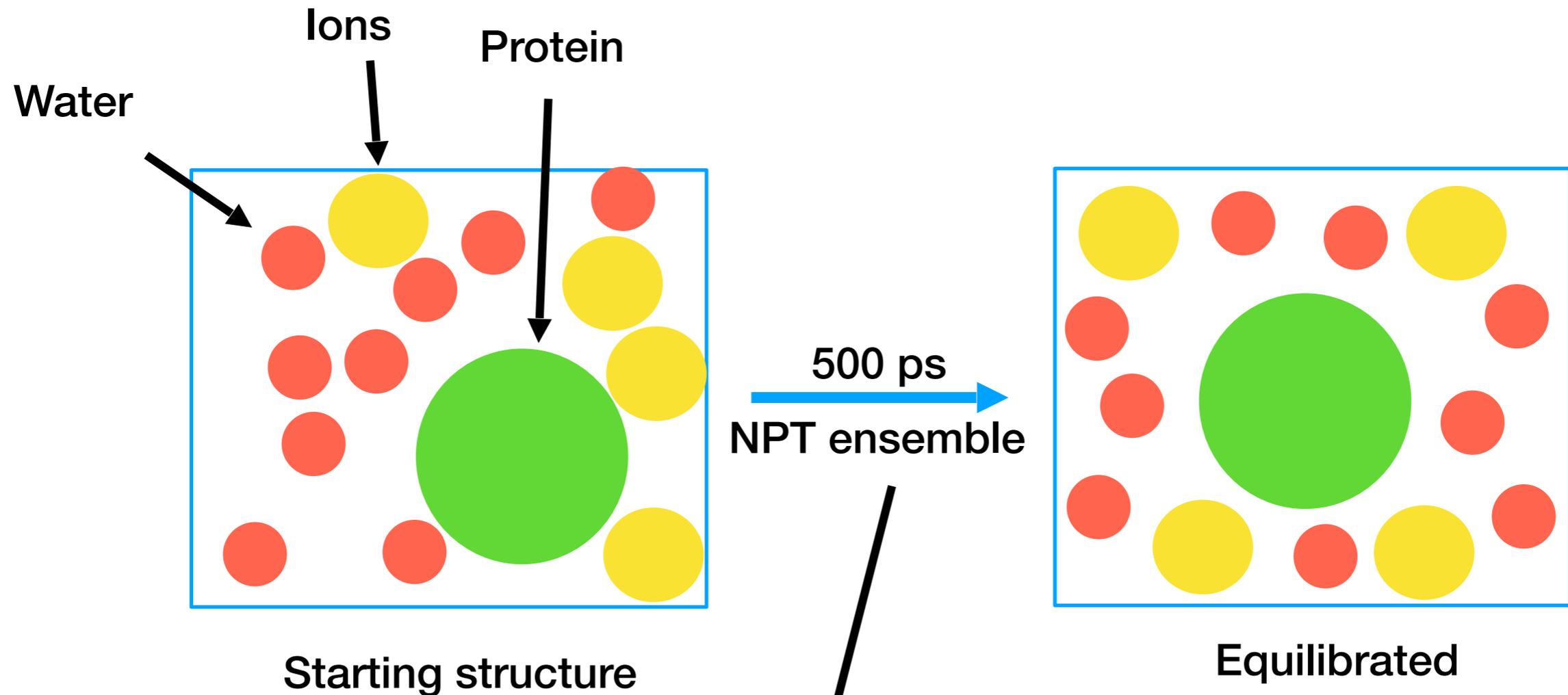
Allows the system to evolve in time indefinitely, that system will eventually pass through all possible states.

experimentally relevant information concerning structural, dynamic and thermodynamic properties may then be calculated using a feasible amount of computer resources.

Energy minimization

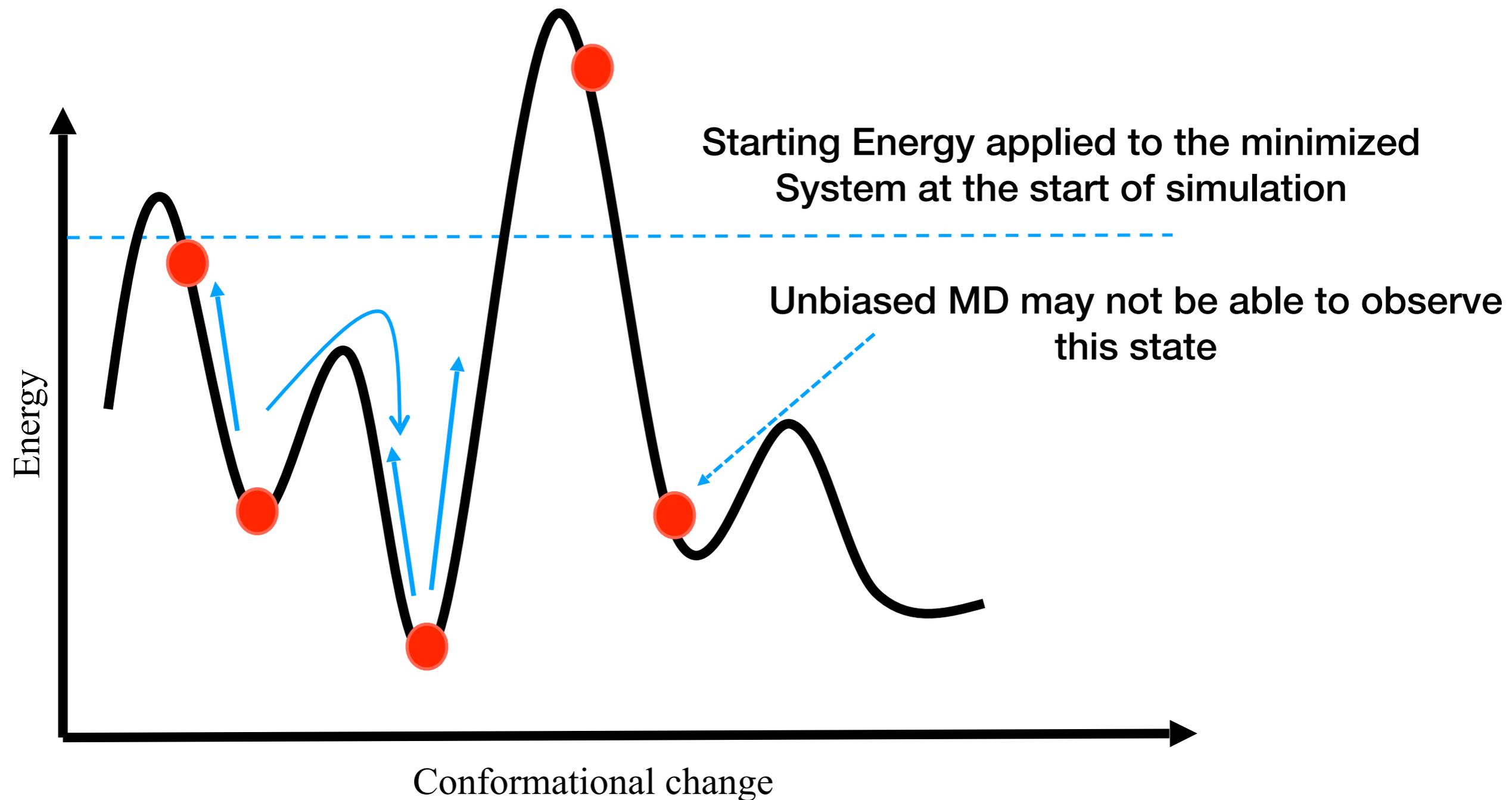


Equilibration

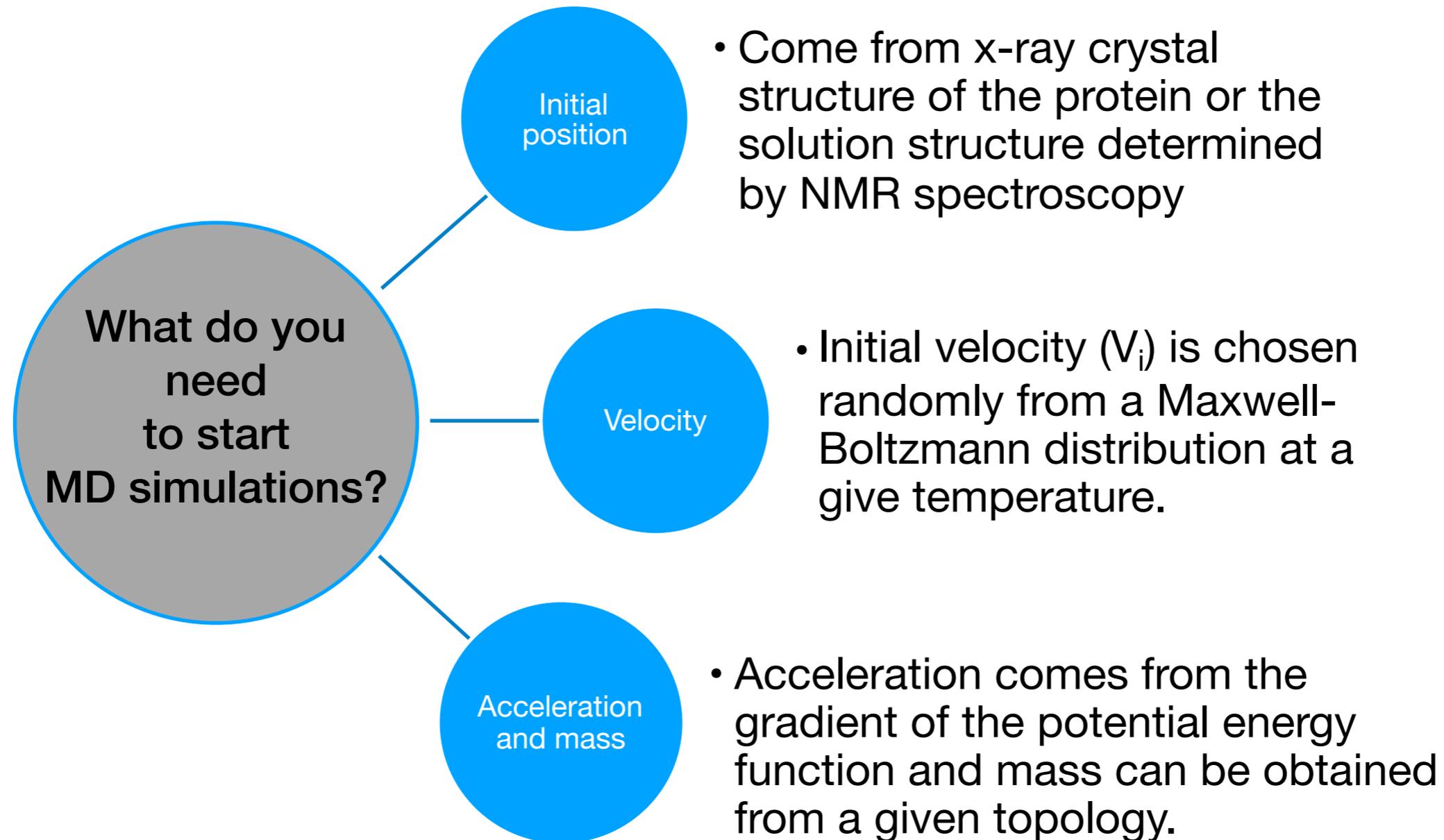


We need Thermostat to control temperature and pressure around the constant level

MD: Change in conformation over time using a force field



MD simulations ingredients



Atomic positions

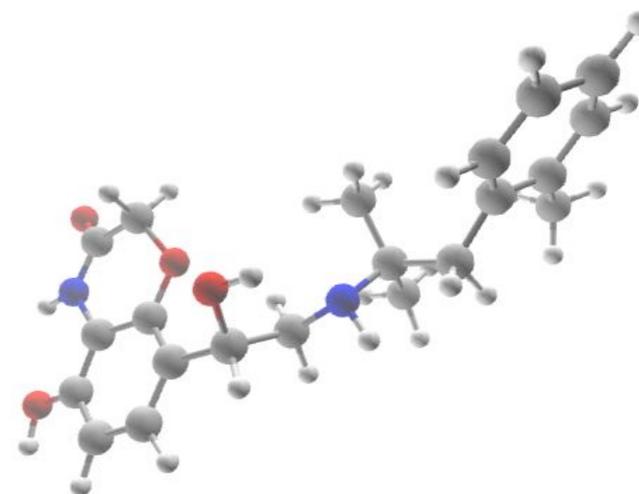
						X	Y	Z
ATOM	1	N	GLU	A	30	62.650	49.480	18.010
ATOM	2	H1	GLU	A	30	62.060	48.690	18.210
ATOM	3	H2	GLU	A	30	62.440	49.710	17.050
ATOM	4	H3	GLU	A	30	63.650	49.370	18.120
ATOM	5	CA	GLU	A	30	62.100	50.540	18.870
ATOM	6	HA	GLU	A	30	62.420	51.530	18.550
ATOM	7	CB	GLU	A	30	60.580	50.580	18.870
ATOM	8	HB1	GLU	A	30	60.160	51.180	19.670
ATOM	9	HB2	GLU	A	30	60.300	49.540	19.010
ATOM	10	CG	GLU	A	30	59.930	50.950	17.530
ATOM	11	HG1	GLU	A	30	60.200	50.260	16.730
ATOM	12	HG2	GLU	A	30	60.360	51.910	17.220
ATOM	13	CD	GLU	A	30	58.420	51.060	17.580
ATOM	14	OE1	GLU	A	30	57.810	51.260	16.500
ATOM	15	OE2	GLU	A	30	57.770	50.760	18.610
ATOM	16	C	GLU	A	30	62.600	50.340	20.300
ATOM	17	O	GLU	A	30	62.390	49.270	20.860

Topology

RESI	p0g	GROUP	Atom type	CHARGE
ATOM	C1	CG331	-0.267	
ATOM	C2	CG331	-0.272	
ATOM	C3	CG331	-0.272	
ATOM	C7	CG2R61	-0.115	
ATOM	C8	CG2R61	-0.108	
ATOM	C9	CG2R61	-0.109	
ATOM	C10	CG2R61	-0.117	
ATOM	C11	CG2R61	-0.113	
ATOM	C12	CG2R61	-0.108	
ATOM	C13	CG321	-0.004	
ATOM	C14	CG324	0.187	
ATOM	C15	CG321	-0.179	
ATOM	C19	CG201	0.501	
ATOM	C20	CG2R61	-0.004	
ATOM	C21	CG2R61	0.107	
ATOM	C22	CG2R61	-0.011	
ATOM	C23	CG2R61	0.003	
ATOM	C24	CG2R61	0.150	
ATOM	C25	CG2R61	0.154	
ATOM	C26	CG311	0.132	
ATOM	C27	CG301	0.366	
ATOM	N16	NG3P2	-0.392	
ATOM	N17	NG2S1	-0.465	
ATOM	O4	OG2D1	-0.524	
ATOM	O5	OG311	-0.532	
ATOM	O6	OG311	-0.636	
ATOM	O18	OG3R60	-0.312	
ATOM	H11	HGA3	0.090	

BOND	H1	C11
BOND	H5	O5
BOND	O5	C21
BOND	C11	C21
BOND	C11	C12
BOND	H2	C12
BOND	C21	C24
BOND	C12	C23
BOND	H6	O6
BOND	O6	C26
BOND	C24	N17
BOND	C24	C25
BOND	C23	C25
BOND	C23	C26
BOND	H17	N17
BOND	N17	C19
BOND	C25	O18
BOND	C26	H26
BOND	C26	C14
BOND	H42	C14
BOND	C19	O4
BOND	C19	C13
BOND	C14	H41
BOND	C14	N16

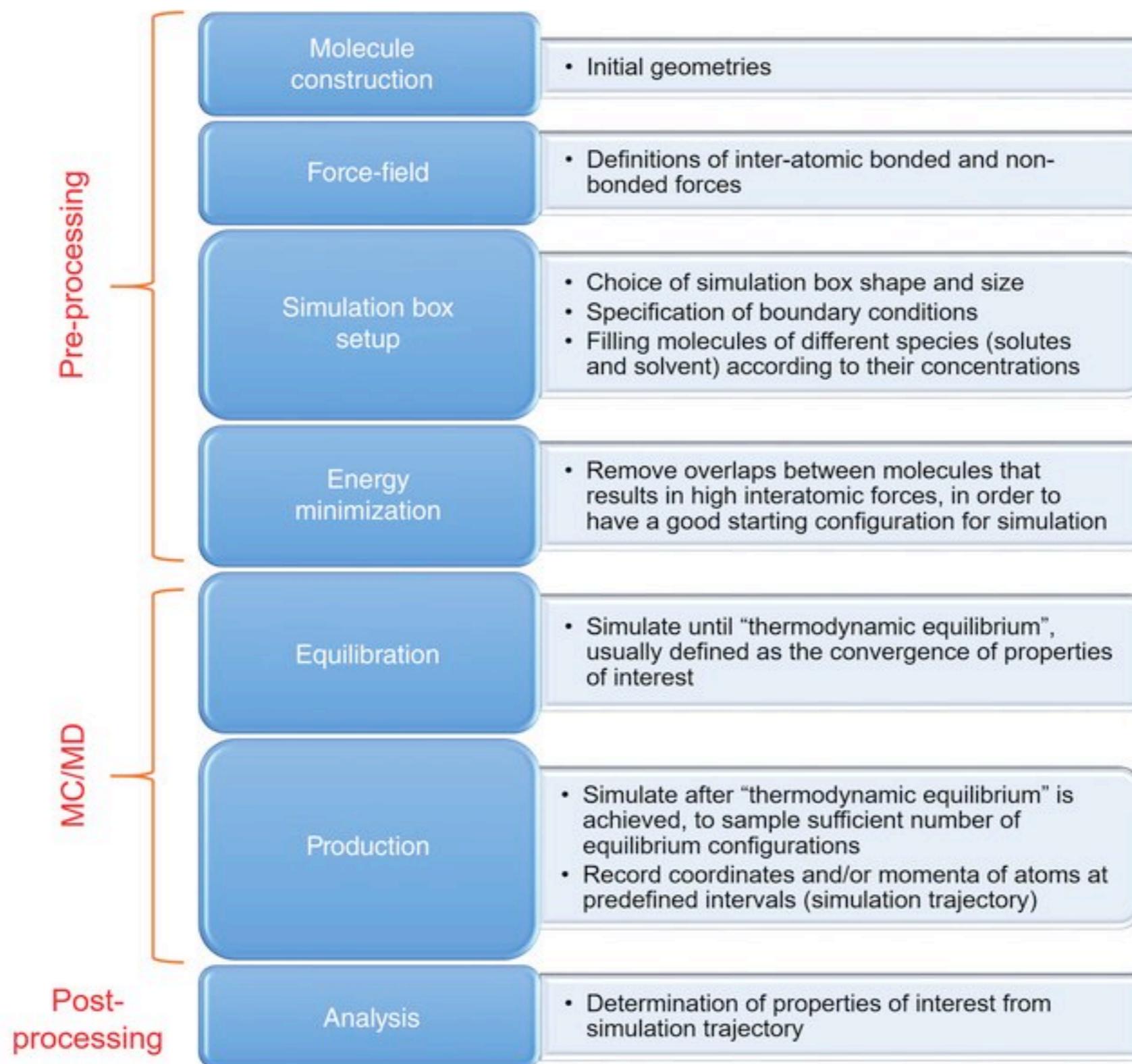
How atoms are connected to each other



Parameter

BONDS						
CG301	NG3P2	200.00	1.4900			
ANGLES						
CG321	CG301	NG3P2	40.00	110.00		
CG331	CG301	NG3P2	40.00	110.00		
CG2R61	CG311	CG324	51.80	107.50		
CG2R61	CG311	OG311	75.70	110.10		
CG201	CG321	OG3R60	112.00	111.00		
CG2R61	CG321	CG301	51.80	107.50		
CG301	NG3P2	CG324	40.00	115.20		
CG301	NG3P2	HGP2	30.00	110.80		
CG2R61	OG3R60	CG321	20.00	99.00		
DIHEDRALS						
NG2S1	CG201	CG321	OG3R60	0.4000	1	0.00
OG2D1	CG201	CG321	OG3R60	0.0000	1	0.00
CG311	CG2R61	CG2R61	OG3R60	2.4000	2	180.00
CG321	CG2R61	CG2R61	CG331	2.4000	2	180.00
NG2S1	CG2R61	CG2R61	OG3R60	2.4000	2	180.00
CG2R61	CG2R61	CG311	CG324	0.2300	2	180.00
CG2R61	CG2R61	CG311	OG311	0.0000	2	0.00
CG2R61	CG2R61	CG321	CG301	0.2300	2	180.00
CG2R61	CG2R61	OG3R60	CG321	1.6200	2	180.00
CG2R61	CG2R61	OG3R60	CG321	0.1900	4	180.00
CG331	CG301	CG321	CG2R61	0.0400	3	0.00
NG3P2	CG301	CG321	CG2R61	0.2000	3	0.00
NG3P2	CG301	CG321	HGA2	0.1950	3	0.00
NG3P2	CG301	CG331	HGA3	0.2000	3	0.00
CG321	CG301	NG3P2	CG324	0.1000	3	0.00
CG321	CG301	NG3P2	HGP2	0.1000	3	0.00
CG331	CG301	NG3P2	CG324	0.1000	3	0.00
CG331	CG301	NG3P2	HGP2	0.1000	3	0.00
CG2R61	CG311	CG324	NG3P2	0.2000	3	0.00
CG2R61	CG311	CG324	HGA2	0.0000	3	0.00
OG311	CG311	CG324	NG3P2	0.1950	3	0.00
CG2R61	CG311	OG311	HGP1	2.1000	1	0.00
CG2R61	CG311	OG311	HGP1	1.4000	2	0.00
CG2R61	CG311	OG311	HGP1	0.7400	3	0.00
CG201	CG321	OG3R60	CG2R61	0.7000	3	0.00
HGA2	CG321	OG3R60	CG2R61	0.9000	3	0.00
CG311	CG324	NG3P2	CG301	0.4000	1	0.00
CG311	CG324	NG3P2	CG301	0.2500	2	0.00
CG311	CG324	NG3P2	CG301	0.6000	3	0.00
HGA2	CG324	NG3P2	CG301	0.1000	3	0.00

Scheme of MD simulations



Molecular simulations in drug delivery: Opportunities and challenges, Ratna S. Katiyar Prateek K. Jha, 2019, Computational Molecular Science.